

Omega-3 fatty acids, lipids and apoE lipitation in Alzheimer's disease: a rationale for multi-nutrient dementia prevention

Marcus O.W. Grimm^{1,2,3*#}, Daniel M. Michaelson^{4#}, Tobias Hartmann^{1,2,3*#}

¹Experimental Neurology, Saarland University, Kirrberger Str. 1, 66421 Homburg/Saar, Germany;

²Neurodegeneration and Neurobiology, Saarland University, Kirrberger Str. 1, 66421 Homburg/Saar,

Germany; ³Deutsches Institut für DemenzPrävention (DIDP), Saarland University, Kirrberger Str. 1,

66421 Homburg/Saar, Germany; ⁴The Department of Neurobiology, The George S. Wise Faculty of

Life Sciences, The Sagol School of Neuroscience, Tel Aviv University, Israel

* correspondence should be addressed to:

Marcus O.W. Grimm (Kirrberger Str. 1, Gebäude 61.4, 66421 Homburg, Germany; phone: +49-6841-1647927; fax: +49-6841-1647801; marcus.grimm@uks.eu)

Tobias Hartmann (Kirrberger Str. 1, Gebäude 61.4, 66421 Homburg, Germany; phone: +49-6841-1647918; tobias.hartmann@mx.uni-saarland.de)

all authors contributed equally

Running title: lipids, apoE and multi-nutrients in Alzheimer prevention

Abbreviations:

A β , amyloid- β ; ADAM, a disintegrin and metalloprotease; AICD, APP intracellular domain; AD, Alzheimer's disease; APP, amyloid precursor protein; BACE, β -secretase APP cleaving enzyme; CSF, cerebrospinal fluid; EPA, eicosapentaenoic acid; FAD, familial Alzheimer's disease; FC, fortasyn; GCS, glucosylceramide-synthase; GWAS, genome wide association study; HMGCR, 3-

hydroxy-3-methyl-glutaryl-CoA-reductase; I-CLIPs, intramembrane cleaving proteases; IDE, insulin-degrading enzyme; LDL, low density lipoprotein; MCI, mild cognitive impairment; NPD1, neuroprotection D1; PE, phosphatidyl-ethanolamine; PS, presenilin; ROS, reactive oxidative species; sAPP α , α -secreted APP; sAPP β , β -secreted APP; SPT, serin-palmitoyl transferase; SIP, sphingosine-1-phosphat

Abstract

In the last decade it has become obvious that Alzheimer's disease (AD) is closely linked to changes in lipids or lipid metabolism. One of the main pathological hallmarks of AD is amyloid- β (A β) deposition. A β is derived from sequential proteolytic processing of the amyloid precursor protein (APP). Interestingly, both, the APP and all APP secretases are transmembrane proteins which cleave APP close to and in the lipid bilayer. Moreover, apolipoprotein E4 (apoE4) has been identified as the most prevalent genetic risk factor for AD. ApoE is the main lipoprotein in the brain, which has an abundant role in the transport of lipids and brain lipid metabolism. Several lipidomic approaches revealed changes in the lipid levels of CSF or in *post mortem* AD brains. Here we review the impact of apoE and lipids in AD focusing on the major brain lipid classes, sphingomyelin, plasmalogens, gangliosides, sulfatides, docosahexaenoic acid and eicosapentaenoic acid as well as on lipid signaling molecules like ceramide and sphingosine-1-phosphat. As nutritional approaches showed limited beneficial effects in clinical studies, the opportunities of combining different supplements in multinutritional approaches are discussed and summarized.

Key words:

Alzheimer's disease, apolipoproteins, lipids, nutrition, sphingomyelin, ceramide, DHA, amyloid- β ; multi-nutrients, n3 fatty acids

Introduction

Alzheimer's disease (AD) is a progressive irreversible neurodegenerative disorder and the most common cause of dementia in the elderly, currently affecting approximately 35 million worldwide (1). Clinically AD is characterized by a loss of short-term memory, intellectual performance and disorientation with a complete loss of memory and mental functions in advanced stages of the disease (2).

The first notion on a possible involvement of lipids in AD dates back to Alois Alzheimer. By examining the brain of Auguste Deter Alzheimer noticed, besides the infamous fibrillary and plaque pathology, lipid granule accumulation in the glia (3, 4). Almost a century later Roses reported that the AD risk increased dramatically with increasing number of apolipoprotein E4 (apoE4) alleles in families with late onset AD (5), a finding immediately confirmed by several other groups (5-8). Among others, Sparks *et al.* showed that cholesterol might represent a molecular risk factor for AD reporting that cholesterol fed to rabbits increased cerebral A β deposition (9), a finding that was later successfully reproduced in AD transgenic mouse models (10, 11). Beyreuther and colleagues found that cholesterol increases neuronal A β generation (12), which could be reversed by cholesterol depletion (13) and treatment with the cholesterol lowering drug simvastatin successfully lowered cerebral A β levels, including the relevant A β 42 species (14) in guinea pigs. At the same time epidemiological evidence supported the presence of an AD – cholesterol – statin link (15). Cholesterol was identified as an early risk factor for AD (16), indicating that cholesterol plays an important role approximately at the same time when amyloid deposition initiates. Clinical trials however found no benefit of atorvastatin or simvastatin treatment in mild to moderate AD (17, 18).

ApoE

Apolipoprotein E (apoE) exists as three different isoforms termed apoE2 apoE3 and apoE4 of which apoE4 is the most prevalent genetic risk factor of AD (5, 19, 20). ApoE is the main lipoprotein in the

brain and plays an important role in intercellular transport of lipids and in brain lipid metabolism. The pivotal role of lipid related mechanisms in the normal physiological function of apoE led to the suggestion that the pathological effects of apoE4 are driven by a lipid related mechanism. This hypothesis is further supported by recent genome-wide association (GWA) studies that identified genes involved in cholesterol metabolism or transport as AD susceptibility genes (21, 22). GWA analysis identified two novel loci, clusterin (CLU), also known as ApoJ, on chromosome 8 and phosphatidylinositol binding clathrin assembly protein (PICALM) on chromosome 11 (21). CLU is a heterodimeric molecular chaperone involved in protein folding of secreted proteins and has been found to interact with the soluble form of A β (23, 24), whereas PICALM is important for clathrin-mediated endocytosis, an essential step in the intracellular trafficking of proteins and lipids. The evidence that support this hypothesis and the mechanisms underlying the role of lipids in mediating the pathological effects of apoE4 will be presently discussed.

ApoE4 and lipids in AD

ApoE4 inhibits serum lipolysis leading to decreased delivery of free fatty acids (FAs) into the brain and their incorporation into glia and neurons (25, 26). Accordingly, a diet enriched with the omega-3 fatty acid (n3-FA) which is beneficial to non apoE4 AD subjects was found to be ineffective in the corresponding apoE4 subjects (27). Serum cholesterol levels are elevated in apoE4 carriers (28). Since elevated serum cholesterol levels at mid-life are associated with increased AD risk (29, 30), it has been suggested that serum cholesterol may play a role in mediating the lipid related pathological effects of apoE4. Corresponding measurements of the total cholesterol and phospholipid levels in the cerebrospinal fluid (CSF) revealed that they are not affected by apoE4 (31). However, further studies that focused on distinct glycerolipids revealed specific effects of apoE4. Accordingly, the levels of sphingosin-1-phosphate in the hippocampus are specifically lower in apoE4 positive AD patients than in corresponding non-apoE4 carriers (32) and the levels of phosphoinositol bisphosphate are reduced in *post mortem* brain tissue of apoE4 carrier as well as in brains of corresponding apoE4 targeted replacement (TR) mice (33).

TR mice in which the endogenous mouse apoE has been replaced by either human apoE4 or human apoE3 have been used extensively to study the mechanisms underlying the pathophysiological effects of apoE4 and the role of lipids in mediating them. Measurements of the effects of apoE2, apoE3 and apoE4 on the brain and plasma levels of glycerolipids and cholesterol did not however reveal a pronounced effect of the apoE genotype on these lipids (34, 35). Nonetheless there was a pronounced effect of diet on the magnitude of the apoE4 phenotype such that the brain pathological phenotype and cognitive effects of apoE4 were reduced by high DHA diet and accentuated by a high cholesterol (34, 36) and carbohydrate diets (37). These human and animal model studies support an involvement of lipids in the pathological effects of apoE4.

Lipidation of apoE4

The lipid driven effects of apoE4 could be due to effects of the apoE genotype on the size, chemical composition and structure of the apoE particles and/or to a lipid related target with which apoE interacts. In the following we discuss findings that show that the apoE isoforms affect differentially the size and extent of lipidation of the apoE particles. The first indication that apoE4 and apoE3 differ in their lipoprotein related properties was obtained over 20 years ago by Weisgraber and colleagues who showed that following incubation with serum lipoproteins, apoE4 but not apoE3 localizes preferentially with vLDL (38). Separation of human CSF apoE by density gradient ultra-centrifugation revealed that apoE4 carriers have the highest level of lipid depleted apoE particles compared with the apoE2 and apoE3 groups and that apoE2 had the highest levels of highly lipidated particles compared to the other groups (39). The effect of the apoE genotype on the size of the CSF apoE particle was assessed by non-denaturing gel electrophoresis which revealed that the apoE2/E3 subjects had significantly larger apoE particles than apoE3/E3 subjects who had significantly larger apoE complexes than apoE3/E4 and apoE4/E4 individuals (40). Similar results were obtained utilizing apoE4 and apoE3 TR mice in which brain apoE4 is associated with smaller less lipidated particles than apoE3 (40). Consistently with these findings it was shown that viral mediated expression of the different apoE isoforms in apoE TR mice leads in the case of apoE4 to the expression of the smallest

and least lipidated apoE particles (41). Taken together these observations show that brain and CSF apoE4 particles are smaller and less lipidated than those of non apoE4 carriers (41). The mechanisms underlying the isoform specific effects of apoE4 on the size and extent of lipidation of apoE and the role of this effect in mediating the pathological effects of apoE4 are being addressed in the following sections.

ABCA1 and the lipidation of apoE4

The ATP binding cassette transporter A1 (ABCA1) is a transmembrane protein that translocates cholesterol and phospholipids to lipid free lipoproteins leading to the generation of HDL-like particles (42). In the periphery apoA-1 and apoE are the main substrates of ABCA1 whereas in the brain apoE is its main substrate. Binding of the lipid free lipoprotein to ABCA1 is critical for initiation of lipid efflux (43). This is followed by translocation of the lipids from the plasma membrane and intracellular pools to the lipoproteins via ABCA1 and to the subsequent release of the lipidated HDL-like particles into the interstitial fluid. The ABCA1 lipidated particles can be further lipidated by the ABCG1 transporter which, unlike ABCA1, lipidates partially lipidated but not lipid free lipoproteins (44). An important development in the study of ABCA1 was the discovery of mutations in this protein that cause Tangier disease and lead to impaired cellular cholesterol efflux and low levels of HDL particles (42). A recent large scale genetic study established that a well-established loss of function mutation of ABCA1 is strongly associated with a higher risk of AD (45). The importance of ABCA1 for the lipidation of apoE in the brain was also highlighted by experiments utilizing ABCA1 knockout mice. This revealed that ABCA1 deficiency in mice reduced the amount of cholesterol in the CSF and that the corresponding CSF apoE containing particles were smaller and contained less cholesterol and less apoE than those of ABCA1 containing mice (46, 47). Whereas ABCA1 deficiency decreases the level of the apoE protein, the corresponding mRNA were not affected by this treatment suggesting that apoE is stabilized by ABCA1 (46). Interestingly, whereas ABCA1 is needed for the production of normally lipidated brain apoE, apoJ does not depend on ABCA1, consistent with the idea that apoE and apoJ reside on distinct lipoprotein particles (48).

The central role of ABCA1 in the lipidation of apoE led to the examination of the possibility that the hypolipidation of apoE4 could be reversed by pharmacological activation of ABCA1, this was first examined by treating apoE3 and apoE4 mice with the RXR ligand bexarotene. This treatment increased the level of ABCA1 in the brains of the apoE4 and apoE3 mice and was associated with reversal of the hypolipidation of apoE4 but with no change in the lipidation of apoE3 (49). Similar results were obtained utilizing the peptide CS-6253 (50) which was previously shown to directly activate ABCA1 *in vitro* (51). Bexarotene activates ABCA1 by increasing the expression of the ABCA1 and subsequently increasing the levels of the ABCA1 protein whereas the effects of CS-6253 are mainly due to activation of ABCA1 driven lipid efflux. Although the molecular mechanisms underlying these processes are not fully worked out yet, the joint take home message is that the hypolipidation of brain apoE4 can be counteracted by activation of brain ABCA1 activity.

Measurement of the effects of the apoE genotype on apoE serum levels of the apoE4 and apoE3 mice revealed that, like in the brain, the levels of apoE4 in the serum are lower than those of apoE3 and that they elute faster following gel permeation chromatography (49). Activation of ABCA1 reversed the elution profile of apoE4 and rendered it more similar to that of serum apoE3, which was not affected by this treatment (49).

The role of hypolipidation in mediating the pathological effects of apoE4

This has been investigated by two complimentary approaches. The first focuses on the general *in vivo* neuronal and behavioral phenotype of apoE4 and on determination of the extent to which it can be counteracted by reversal of the hypolipidation of apoE4. The second approach focuses on specific apriori apoE4 driven mechanisms such as cross talk with A β and synaptic impairments.

Counter acting the brain and cognitive effects of apoE4 by reversal of its hypolipidation: nd Treatment of the apoE4 and apoE3 TR mice with either bexarotene or the ABCA1 agonist CS-6253, both of which reverse the hypolipidation of apoE4 (49), reversed the apoE4 driven accumulation of A β 42 and hyperphosphorylated tau in hippocampal neurons as well as the associated synaptic impairment and cognitive deficits of these mice (49). These findings suggest that hypolipidation plays a pivotal role in

mediating the pathological effects of apoE4 and point at ABCA1 as a promising therapeutic target. Beneficial effects of ABCA1 activation were also observed in the PDAPP mouse model where amyloid deposition was reduced by overexpression of ABCA1 (52). Cell culture studies revealed that neurite outgrowth is stimulated by PUFA containing phospholipids suggesting that PUFA play an important role in mediating the lipidation dependent effects of apoE (53). As the phenotype of the apoE4 mice is complex and involves the AD molecules Tau and A β as well as synaptic molecules and apoE receptors, additional more targeted models are needed for assessing the cellular mechanisms and processes which drive the apoE4 hypolipidation related effects of apoE4.

ApoE4 lipidation and A β : *In vitro* binding studies revealed that human apoE binds to A β isoform specifically in a lipid dependent fashion. Accordingly lipidated apoE3 binds to A β more avidly than the corresponding apoE4. The affinity of both apoE isoforms to A β and its apoE isoform specificity are both reduced following delipidation of apoE (54, 55). The finding that lipidated apoE3 binds more avidly to A β than apoE4 led to the suggestion that apoE3 is involved in the clearance of A β from the central nervous system and that this physiological mechanism is impaired in apoE4. This was confirmed by *in vivo* studies with mice which express the different human apoE isoforms together with the A β precursor APP, which revealed that the $t_{1/2}$ of A β in the brain is longer in the apoE4 than the apoE3 mice and that, like in AD, this was associated with increased deposition of A β in the apoE4 mice (56-58). In view of these findings it is possible that the accumulation of A β in hippocampal neurons of the apoE4 mice (49, 59) is due to the hypolipidation of apoE4 and the resulting impaired clearance of A β . This suggests that dislipidation plays an important role in mediating the A β dependent apoE4 driven pathological effects. Although the *in vitro* data suggest that these effects are driven by direct interactions between apoE4 and A β , recent *in vitro* findings suggest that apoE4 and A β interact minimally under physiological conditions and that consequently indirect mechanisms may play a role in mediating the cross talk between apoE and A β (57).

ApoE4 lipidation and synaptic pathology: Neurite outgrowth requires a supply of lipids for membrane expansion. Indeed neurite outgrowth by numerous cell types, including astrocytes and neurons, was activated following incubation with lipidated apoE3 whereas incubation with apoE4 either inhibits or has no effect on this process (60-63). However as recombinant non lipidated apoE4 has the same effect

as lipidated apoE4 (61), it is possible that these neurite related effects of apoE4 are also driven by a lipid independent mechanism. Several other mechanisms are possible, apoE4 has been implicated, in addition to A β and synaptic mechanism, with numerous other processes. These include apoE receptors, inflammation, apoE4 degradation and the vascular system (64). The roles of apoE4 hypolipidation in mediating these apoE4 phenotypes remain to be determined.

Amyloid precursor protein (APP) processing

Main pathological hallmarks of AD are extracellular amyloid plaques composed of aggregated amyloid- β (A β) peptides (65, 66) and intracellular neurofibrillary tangles due to hyperphosphorylation of the microtubule-associated protein tau (67, 68). A β peptides are generated by sequential proteolytic processing of the amyloid precursor protein (APP), a large type I transmembrane protein (69). The amyloidogenic processing of APP is initiated by β -secretase BACE 1 (70, 71) which cleaves APP within its extracellular/intraluminal N-terminal domain, shedding off β -secreted APP (sAPP β). The remaining membrane-tethered C-terminal fragment (β CTF/C99) is further cleaved by γ -secretase, to release A β peptides in the extracellular space (72, 73) and the APP intracellular domain (AICD) in the cytosol which is discussed to translocate to the nucleus and to regulate gene transcription of different target genes (74-83). A β peptides vary in length from 37 to 43 amino acids with the main products being A β 38, A β 40 and A β 42 (73, 84-87). Although A β 42 only accounts for approximately 10% of total secreted A β peptides, the A β 42 species represents the major component of amyloid plaques. Due to the two additional amino acids isoleucine and alanine, A β 42 is more hydrophobic and aggregates faster than A β 40 peptides (88, 89). Before manifested as amyloid plaques in vulnerable brain regions like hippocampus and cortex these peptides form oligomers, protofibrils and fibrils. Small oligomers of A β up to 50 A β subunits are discussed to be the most toxic variant of A β (90-93).

APP secretases

The γ -secretase which generates the C-terminus of A β peptides is a heterotetrameric protein complex consisting of Presenilin 1 or 2 (PS1 or PS2), nicastrin, anterior-pharynx defective 1 a or b and presenilin enhancer 2 (94). PS1 or PS2 represent the catalytically active components of the γ -secretase complex (95, 96). Mutations in the presenilins cause familial early onset Alzheimer's disease caused by an increase in A β 42 peptides and a decrease in A β 40 (97-100). The γ -secretase complex belongs to the intramembrane cleaving proteases (I-CLIPs) with the peculiar property to cleave transmembrane proteins within their hydrophobic transmembrane domains. I-CLIPs including γ -secretase are themselves membrane-embedded proteases with multiple transmembrane domains, emphasizing the importance of the lipid microenvironment of the membrane for proper function. Indeed, amyloidogenic processing of APP by β - and γ -secretase is discussed to occur within lipid rafts (101-104), membrane microdomains enriched in cholesterol and sphingolipids (13, 105, 106). The importance of lipids for AD is further strengthened by observations that several lipid classes can influence proteolytic processing of APP, including cholesterol, sphingolipids, polyunsaturated fatty acids (PUFAs), plasmalogens, *trans*-FAs and phytosterols (13, 107-117) and that several lipid classes have been found to be altered in AD *post mortem* brain (118-121). Alternatively to the initial cleavage by β -secretase, APP can be cleaved within its N-terminal domain by α -secretases. The α -secretases have been identified as members of the ADAM family (a disintegrin and metalloprotease), they cleave APP within the A β domain, thus preventing the generation of A β peptides (122-126). Cleavage of the remaining C-terminal fragment α -CTF/C83 by the γ -secretase complex liberates non-toxic p3 peptides (127). Interestingly the α -secretases as well as BACE1 are also membrane-embedded proteases. In contrast to β -secretase processing of APP which is discussed to be lipid raft associated, α -secretase cleavage of APP seems to take place outside of lipid rafts (128, 129). APP processing into A β , AICD and other APP fragments is a key part of the physiological function of APP in lipid sensing and lipid regulation (76, 77, 130, 131), which provides a rationale why APP cleavage is very sensitive to alterations in the membrane lipid composition (78, 111, 132). Beside a possible direct effect of lipids on APP processing, lipids might be important regulators for lateral movement of proteins within the phospholipid bilayer and are therefore critical for substrate/enzyme interaction and thus a correct balance between amyloidogenic and non-amyloidogenic APP processing (133). Moreover, disturbing

or changing the lipid composition of the membrane might therefore play a crucial role in the pathogenesis and treatment of AD. The following text provides a brief overview of the role of the major lipids in AD.

Sphingomyelin

Sphingomyelin (SM) is a major constituent of cellular membranes and is one of the major lipids in the human brain. The structures of the discussed sphingolipids, which are changed in AD, are shown in figure 1. It occurs in high concentrations in neuronal membranes and in the myelin sheets. Besides its role as a signaling molecule it has a crucial function in the structure of membranes. Due to its highly intermolecular interactions, which are mediated by the 2-amide group, 3-hydroxy group and partially by the 4,5-*trans* double bond of the sphingoid-base, SM has a crucial function in forming membrane microdomains. These microdomains, also called lipid rafts, are signalling platforms and are discussed to include the secretases involved in the amyloidogenic pathway (13, 101-104). In *post mortem* brains of patients suffering from AD a reduction in SM and an elevation of ceramide has been reported compared to age-matched control brains (134). Interestingly SM is a precursor for ceramide, a reaction which is catalyzed by the action of the sphingomyelinases (SMases). Acid SMase and acid ceramidase expression was accompanied by the observed change in the lipid pattern (134). Mechanistically it has been shown that A β , especially A β 42, can directly activate SMases in the picomolar range. In return, inhibition of SMases leading to an increased SM level resulted in a decrease of A β production (131). In line, mutations in PS1 causing familial Alzheimer's disease (FAD) showed increased SMase activity (131). Under pathological conditions high A β 42 concentrations resulting in oligomers, protofibrils and fibrils and leading to oxidative stress and inflammatory mechanisms have been demonstrated to additionally activate nSMase (135, 136). Similar results have been found by Jana *et al.* showing that fibrillary A β peptides activate nSMases via NADPH oxidase mediated mechanisms finally leading to the loss of neurons (137, 138). A more recent paper showed a significantly increased nSMase activity not only in human *post mortem* brains but also in plasma and fibroblasts derived from AD patients. Notably no elevation was found in samples from Parkinson disease patients indicating that increased

aSMase activity is a specific signature in AD (139). Elevated aSMase activity was also found in a transgenic AD mouse model consisting of mutated APP and PS1; especially in neurons and brain but also in plasma and fibroblasts these transgenic mice showed an increased aSMase activity (139). Combining this transgenic mouse model with an aSMase +/- mouse revealed a significantly reduced A β deposition and memory dysfunction (139). Recently a convergent study was published; nSMase 2 deficient mice combined with 5xFAD mice revealed improved cognition and showed an ameliorated AD pathology. Total A β 42 and plaque burden but also additionally Tau phosphorylation was decreased and the transgenic mice showed an improved cognition in fear-conditioned learning tasks (140). Another connection between AD, SM and SMases can be indirectly drawn from the fact that SM is decreased in depression accompanied by an increase in aSMase activity (141). A quantitative meta-analysis revealed that depression is a major risk factor for incidence of dementia including AD and mild cognitive impairment (MCI) (142).

Analysing different metabolites in biofluids of AD patients revealed that in plasma two ceramide species were increased and eight SM species were decreased in AD, accompanied by a total increase in ceramides, both in serum and CSF (143). Beside these unambiguous results it should be mentioned that in case of biomarkers the role of SM is not completely understood and controversial results are reported in literature. For example Koal *et al.* report an increased SM species (SM(d18:1/18:0)) to be significantly enhanced in CSF samples of AD patients and are suggested as a biomarker for AD with a specificity of 76% and a sensitivity of 66% with a cut-off of 546nM (144). Interestingly this study also found an increased acyl-carnitine (C3-DC-M/C5-OH). Acyl-carnitines occur under physiological conditions in the cytosol and are used to transport FAs in the mitochondria by the carnitine-carrier-system. An enhanced level of acyl-carnitines in CSF or in the extracellular space might indicate neuronal loss or cell death and might help to explain this finding (134). However, elevated SM levels have also been found in prodromal, mild or moderate AD compared to normal cognitive controls suggesting that alterations in SM homeostasis may contribute to early AD pathological processes and are not only due to neuronal loss (145). Another study showed that, in contrast, many other sphingolipids including the sphingomyelins SM39:1, SM41:1 and SM42:1 are decreased. However, this study clearly points out that SM alone is not sufficient as a biomarker and has to be accompanied

by other lipids to increase specificity and sensitivity (146). Utilizing APP/PS1 transgenic mice another study underlines the importance of SM in AD pathology but also points out that alterations might be region specific, which might further help to explain different results in the literature (147). Summing it up, under physiological conditions A β increases SMase activity. This increase is further enhanced under pathological conditions involving reactive oxidative species (ROS) and inflammatory processes. Consequently SM is found to be reduced at least in some specific regions of AD brain. In return SM decreases A β production suggesting that a decreased SM level is not only a consequence but also a cause in AD. In respect to biomarkers the role of SM is not completely understood. Divergent results occur in literature depending on the stage of disease or the biofluid which is analyzed.

Ceramide

The role of ceramide in cellular signaling processes including cell differentiation, proliferation and programmed cell death is well known. Beside the above mentioned SMases, ceramide is also produced by *de novo* synthesis of the serine-palmitoyl transferase (SPT) and some additional enzymes leading to dehydro-ceramide and ceramide in the endoplasmatic reticulum. The concentration of ceramide is tightly linked to the action of the SMases which are already reviewed above. Therefore it is not surprising that most studies found an increased ceramide level associated with AD (134, 148-150). Shot gun lipidomics approaches of early AD stages identified two ceramide species to be significantly lower, additionally five other species trend to be decreased in plasma samples (151). Similar results were found by Mielke *et al.* reporting that high baseline ceramides were associated with an increased risk of AD (152). Interestingly a reduction in Ceramide synthase 2 activity in Braak stage 1,2 in temporal cortex and Braak stage 3,4 in hippocampus and frontal cortex was found (153). Combining the facts that ceramide synthase 2 activity is discussed to generate very long chain ceramides and that very long chain ceramides are precursors for sulfatides and galactosylceramides typically found in myelin, these findings suggest a defect in myelin biosynthesis in the preclinical or early to moderate stages of AD (154). Moreover, pathway and network analysis revealed a role of ceramide in both, inflammatory and anti-inflammatory pathways of AD, further illustrating the complex action and

mechanisms mediated by different ceramides (155). Enhanced levels of ceramides have been shown to stabilize β -secretase and therefore increasing $A\beta$ level (156). Ceramides have been shown to cause mitochondrial depolarisation and permeabilisation, cytochrom-C release, Bcl2-depletion and caspase-3 activation, further leading to neuronal death (134). Besides the impact of ceramide in AD and APP processing, APP processing in return regulates ceramide level. It has been shown that the APP intracellular domain (AICD) is a regulator of the expression of the SPT, the committed step reaction in ceramide *de novo* synthesis spanning a complex regulatory cycle (76). Giving a resume the link between ceramides and AD is widespread. It has to be kept in mind that ceramide function is both highly dependent of the cellular localisation of the ceramides and the ceramide species especially on the FA bound at the sn-2 position. Whereas ceramides with very long chain FAs are abundant in myelin and discussed to be „good players“ in respect to AD, other ceramide species trigger inflammatory and apoptotic processes. Therefore especially for ceramides biomarkers have to be carefully chosen focusing not in general on the lipid class, the ceramides in total, but more precisely on the different ceramide species. Further studies are needed to fully understand the role of ceramides in the context of AD.

Sphingosine and Sphingosine-1-phosphate

As mentioned above Ceramidase has been reported to be altered in *post mortem* AD brains (134). Ceramidase catalyses the degradation of ceramide to sphingosine by cleaving of one FA. By the action of sphingosine-kinase sphingosine can be phosphorylated to sphingosine-1-phosphate (S1P). In return, sphingosine-phosphatases can convert S1P to sphingosine. Additionally S1P can be cleaved by S1P-lyase resulting in phosphoethanolamin and hexadecenal which is an irreversible reaction. Especially the ratio of S1P to ceramide is linked to several cellular processes, e.g. apoptosis, Ca homeostasis or inflammation (157). As these biochemical processes are involved in neurodegeneration a crucial role of sphingosine and sphingosine derivatives can be expected in the pathological events of AD. Indeed it has been shown that S1P is able to bind to BACE1 and modulates its activity (158). Treatment of mouse neurons with sphingosine-kinase inhibitors, RNA interference leading to knock-down of

sphingosine-kinase, or overexpression of S1P degrading enzymes resulted in decreased BACE activity and reduced A β production. Another study demonstrates that increased S1P level caused by S1P-lyase deficiency results in the accumulation of APP and APP C-terminal fragments in lysosomal compartments and a reduction of γ -secretase activity (116). Beside its effect on APP processing deficiency of S1P-lyase has been reported to increase hyperphosphorylation of tau (159). However the impact of S1P in AD is not completely understood. On the other hand it has been shown that the S1P/sphingosine ratio is declined with increased Braak stage. Additionally the decrease of S1P was found in brain regions which are mostly affected by AD pathology. Interestingly the authors point out an association of these results with the apoE status. The S1P/sphingosine ratio was 2.5fold higher in hippocampus of apoE2 carriers compared to apoE4 carriers (32). In line with these results S1P enhances neuronal cell survival distressed by glucose deprivation and glucose reload stress (160) mediated by the activation of the S1P1- and S1P3-receptor signalling pathways. Recently Fingolimod, a drug already used in multiple sclerosis which is a functional S1P1 receptor antagonist, was used to treat 5xFAD transgenic mice (161). A decreased plaque density accompanied by a decrease in soluble plus insoluble A β was found in Fingolimod-treated mice. Additionally Fingolimod attenuated GFAP staining and microglia activation (161). In conclusion sphingosine or sphingosine related pathways seem to be a promising and interesting target in AD. Nevertheless it has to be taken into consideration that S1P is tightly regulated in brain homeostasis in a low picomolar range and affects a broad network of cellular processes. Further studies are needed to elucidate the crosstalk between sphingosine or sphingosine derivatives and AD and to clarify the potential benefits in respect to expected side effects. Furthermore, the above discussed relation between apoE and sphingosine homeostasis suggests that not all patients might equally profit by a sphingosine-based therapy.

Glycosylated sphingolipids

By the addition of galactose or glucose to ceramide sulfatides and gangliosides, two other important lipid classes in brain, are generated. In case of sulfatides the cerebrosid-sulfo-transferase adds a sulfate to the galactosyl-ceramide, a reaction which can be reverted by the arylsulfatase A. Sulfatides

are both present in the white matter, in the myelin but also in membranes of neurons, especially in the axon structure (162). For both, in grey matter and white matter of *post mortem* brains from AD subjects with mild dementia, a decrease of sulfatides has been reported (163). Another study confirmed this finding and revealed that brain samples from subjects with Parkinson disease and in dementia with Lewy Bodies has no changes or even higher sulfatide levels compared to control samples suggesting that sulfatide reduction is a specific event in AD. Additionally the authors suggest that A β accumulation is not a factor directly contributing to a decreased sulfatide level in AD (164). A decreased sulfatide level could also be confirmed in transgenic AD mouse models. Interestingly apoE was identified to mediate sulfatide reduction in transgenic mouse models of AD (165) in an apoE isoform dependent manner. Therefore beside S1P also sulfatides as a second lipid class were identified to crosstalk with apoE-mediated biochemical processes. A more recent study confirmed the reduction of reduced sulfatide levels in preclinical stages of the disease (166).

The glucosylceramide-synthase (GCS) adds glucose to ceramide generating the precursor for the gangliosides. Interestingly it has been reported that GCS is regulated by APP processing (82). Similar results have been found for the GD3-synthase (GD3S), another enzyme in the ganglioside pathway, converting *a*- series to *b*-series gangliosides (78). Importantly, GM3, the substrate for GD3S decreases A β generation whereas the product, GD3, enhances A β production (78). Accordingly, GD3S deficient mice combined with a transgenic AD mouse model, showed a reduced plaque burden and an improvement in the cognitive abilities (129). A more recent study confirming the effect of GD3S deficient mice suggests also an impact of the cholinergic specific ganglioside GT1Aalpha (167) in memory retention. An increase of the β -hexosaminidase which is a catabolic enzyme in ganglioside homeostasis revealed beneficial effects in cognitive tests with transgenic AD mouse models (168). The importance of this catabolic enzyme is also underlined by a study showing a changed hexosaminidase and β -galactosidase in AD patients (169) probably linked with a dysfunction of lysosomal compartments, a known characteristic in AD (170, 171).

Another ganglioside GM1 has been shown to enhance amyloidogenic pathways (172) and additionally gangliosides have been shown to bind A β and therefore influencing the aggregation of A β leading to

oligomers, protofibrils and fibrils (173-178). In line ganglioside-A β complexes have been discussed to build aggregation seeds for oligomers and plaques (108). Not only during aging but also by comparing brain samples of AD patients with age-matched controls revealed a complex change in ganglioside homeostasis. However it has to be pointed out that the heterogeneous results in literature might be explained that changes seem to be brain region specific and differ in the pathological stages of AD (96, 179-182). For ceramides it has already been discussed that the FA also determines the impact of the individual lipid in AD relevant processes (153). Similar results have been found in respect to gangliosides further explaining different results in literature and emphasizing the need of exact and detailed lipidomic analysis including not only the headgroup but also the FA bound at the sn-1 and sn-2 position.

Plasmalogens

Plasmalogens are ether-phospholipids characterized by a vinyl ether bond at the sn-1 position and an ester linkage at the sn-2 position. The chemical structure of the plasmalogens are presented in figure 2. Several studies showed a decrease in phosphatidylethanolamin- and phosphatidylcholin-plasmalogens in AD *post mortem* brains (119, 121, 183). Analysing PS mutation carriers, a study revealed a decrease in two plasmalogen species (PE34:2, PE35:4) in the CSF which correlated with an increased Tau level in CSF and an increased amyloid burden in the brain (184). Plasmalogens are also found to be decreased in erythrocytes of children with Down syndrome. As Down syndrome is a chromosomal aberration on chromosome 21 (Trisomy 21) and APP is located on chromosome 21 (185), these results further strengthen the linkage between APP processing and plasmalogen homeostasis (186). As the vinyl ether is highly susceptible for reactive oxidative species and oxidation, the reduced plasmalogen levels could be caused by the increased oxidative stress occurring during AD pathogenesis. However, we could demonstrate that a reduced plasmalogen level is not only the consequence of pathological changes in the brain, plasmalogen itself is able to reduce γ -secretase activity in neuroblastoma cells and membranes derived from mouse brains (113). These results have been recently confirmed by another group demonstrating that an increased PE-plasmalogen to PE ratio results in an inhibition of γ -

secretase activity (187). Plasmalogens do not only affect APP processing but also in return APP processing tightly regulates plasmalogen homeostasis (77). AICD decreases the expression of the enzyme catalyzing the committed step reaction in plasmalogen synthesis, the alkyl-dihydroxyacetonephosphate-synthase (AGPS), leading to a decreased plasmalogen synthesis. Results were confirmed in several transgenic mouse models lacking APP or only expressing a truncated APP construct devoid of a functional AICD domain (77, 188). These results suggest under physiological conditions a regulatory feed-back cycle in which plasmalogens inhibit APP processing and APP processing in return downregulates plasmalogen *de novo* synthesis. Under pathological conditions this regulatory cycle is affected. Beside the reduction of plasmalogens by ROS, a reduced AGPS protein stability due to peroxisomal dysfunction was reported leading to a reduced plasmalogen level. The reduced plasmalogen level results in an elevated A β production further enhancing oxidative stress and peroxisomal dysfunction. Summing it up the regulatory feed-back cycle, being necessary to adjust a homeostasis, is converted to a feed-forward or futile cycle under pathological conditions enhancing A β production. Because of its potential benefits in AD as a nutritional supplement, efforts have been made to identify sources of plasmalogens in food like marine invertebrates or blue mussels or ascidians and to extract them from these sources (189). Plasmalogens derived from these natural sources have already been shown to suppress activation of caspase-3 in SH-SY5Y cells further implicating a beneficial effect in AD (189). Recently on the 1st International Plasmalogen Symposium in Japan results were presented suggesting beneficial effects of plasmalogens in clinical studies of patients with MCI.

Eicosapentaenoic acid and other polyunsaturated fatty acids

Also in a lower extent than DHA, the n3 PUFA eicosapentaenoic acid (EPA) is present in brain (190). Figure 3 illustrates the structure of DHA and EPA. Mechanistically it has been shown that EPA is able to enhance A β degradation mediated by IDE. Like DHA, EPA is able to bind IDE and directly enhances IDE activity (136). Moreover EPA increases IDE gene expression; in contrast to DHA no alterations between the extracellular and intracellular IDE ratio compared to its corresponding

saturated FA (20:0) was observed (136). It is discussed that EPA affects the gene expression of many inflammation-related genes in immune cells. A recent study revealed that the anti-inflammatory potency of EPA is comparable to DHA (191). Similarly Hopperton *et al.* reported that increasing brain n3-PUFAs decreases some aspects of the inflammatory response to A β (192). In this recent study, brain n3-PUFAs were increased by a transgenic approach using fat-1 transgenic mice, carrying a transgene converting n6- to n3-PUFAs, or dietary means analysing wt littermates which were orally administered to a fish oil or safflower oil diet containing very low levels of n3-PUFAs. At 12 weeks of age intracerebroventricular infusion of A β 1-40 was performed. Compared to wt mice fed with the safflower oil diet, fat1 transgenic mice and wt mice fed with the fish oil diet showed higher levels of brain DHA whereas n6-PUFAs were decreased. Microglia activation was reduced in fat1 transgenic mice at 10 days post-surgery. The wt mice fed with the fish oil diet showed no effect on microglia activation but revealed fewer degenerating neurons.

Additionally n3-FA supplementation was reported to increase A β phagocytosis by macrophages and influences brain amyloidosis. However it has to be mentioned that in this study n3-FAs have been combined with antioxidants and resveratrol, so that no clear mechanistical conclusions to single compounds can be drawn and further experiments are needed preferable with a higher n-number (193). In context of inflammation and PUFAs it should be discussed that effects might also be mediated by dihydroxy- or trihydroxy-metabolites of n3-FAs which are called resolvins. Resolvin D1 was shown to be associated with an increased A β phagocytosis (194) and might also inhibit fibrillar A β induced apoptosis (195). Another enzymatic derivative of n3-FA, neuroprotectin D1 (NPD1), was suggested to downregulate A β production, pro-inflammatory gene expression and promotes cell survival. In the anti-apoptotic bioactivity induced by NPD1, PPAGamma signalling may be involved and a shift from the amyloidogenic to the non-amyloidogenic pathway is discussed to be responsible for the decreased A β production (196). Because of the complex effects of n3 derivatives like neuroprotectin in AD we would like to refer to a previous review in this journal (197).

Although EPA is often combined with DHA in nutritional approaches (198, 199) mechanistical studies about EPA alone and AD related processes are mainly not provided.

Besides EPA PUFAs have been shown to increase non-amyloidogenic processing of APP leading to an increased sAPP α secretion (114, 200, 201). PUFAs have been shown to increase α -secretase activity in neuroblastoma cells and isolated membranes of both, neuronal cell lines and human *post mortem* brains. Interestingly for linolenic acid (18:3) and DHA it was shown that these lipids can increase α -secretase activity utilizing purified ADAM10 enzyme (114). Together with the result that PUFAs are decreased in human *post mortem* AD brains (121, 202), the anti-inflammatory properties, the effect on the non-amyloidogenic pathway and on A β clearance, also other unsaturated FAs than DHA might be interesting targets in AD especially combined with other nutritional approaches. Remarkably, not only the desaturation, the number and position of double-bonds of FAs seem to influence AD relevant mechanisms. Also the conformation is known to have an important impact. Under physiological conditions double-bonds have a *cis*-conformation resulting in an angled, non-linear orientation of the FA in the biomembrane. Especially the resulting angle determines physical properties like membrane fluidity. In contrast *trans*-FAs have a linear orientation and the membrane is more condensed and tightly packed. *Trans*-FAs, which e.g. naturally occur in products of ruminants, enhance amyloidogenic and decrease non-amyloidogenic APP processing by influencing all three secretases (117). Moreover *trans*-FAs seem to alter intracellular APP processing and enhance A β aggregation compared to the corresponding *cis*-conformation (117). In line epidemiological studies revealed that individuals who had an intake of *trans*-FAs of 4.8g/day showed a 5fold higher relative risk to develop AD than subjects consuming 1.8g/day (203) and dietary intake of *trans*-FAs is associated with cognitive decline in the elderly population (204, 205).

Docosahexaenoic acid

As mentioned above not only the headgroup of lipids but also the FAs are crucially linked with AD. DHA (DHA22:6), a polyunsaturated n3-FA, is the most prominent n3-FA in the CNS (206). 30 – 40 % of the FAs in the phospholipids of neuronal plasma membranes and 8% of the human brain dry-weight are lipids containing DHA (206, 207). DHA synthesis is highly limited in humans, therefore the major amount of DHA has to be taken up by diet (208). DHA and other n3-FAs are efficiently transported

across the blood-brain-barrier (209, 210). However a recent study suggests that the apoE4 allele is associated with a decreased transport of DHA to CSF (211). Consequently apoE4, a known risk factor for AD (5, 212-214), is not only related to cholesterol homeostasis and transport but also influences the homeostasis of lipids tightly coupled to AD. As an example we would like again to underline the importance of apoE in sulfatide homeostasis as reviewed above, but also in the transport of PUFAs across the blood-brain-barrier indicated by the recent literature (211, 215).

After being incorporated in phospholipids DHA determines the physical state of membranes such as membrane fluidity (200, 216). Mechanistically it has been shown that DHA increases the non-amyloidogenic processing resulting in an elevated sAPP α (α -secreted APP) level by an increase in ADAM17 protein levels caused by an upregulated gene expression and a decreased protein degradation (201). Additionally DHA attenuates amyloidogenic processing by both affecting β - and γ -secretase activity by independent mechanisms. Whereas BACE1 total protein levels were unchanged, a changed cellular distribution was observed leading to an accumulation of BACE1 at the cell-surface and a decreased intracellular BACE1 level e.g. in the endosomes (201). As BACE1 activity has its optimal activity at an acidic pH-value which is found in endosomes (217-220), the altered localisation of BACE1 might act as an explanation for a decreased BACE1 activity in presence of DHA. For γ -secretase, both, a direct effect of DHA on the enzyme activity was reported, accompanied by a shift of PS1 from the raft microdomains to the non-raft microdomains of the membrane (201). As PS1 contains the active center of the γ -secretase complex (95) the change in PS1 distribution also leads to a shift in γ -secretase activity. Besides the direct effect of DHA on amyloidogenic and non-amyloidogenic pathways we and others could demonstrate that cholesterol *de novo* synthesis especially the activity of the enzyme catalyzing the committed step reaction, the 3-hydroxy-3-methylglutaryl-CoA-reductase (HMGCR), was decreased in presence of DHA (201, 221). Cholesterol is known to increase amyloidogenic and to decrease non-amyloidogenic APP processing (13, 107, 112). Therefore a reduction in cellular cholesterol levels also contribute to the pleiotropic DHA-mediated effects which all synergistically result in a decreased A β level. A β level are not only dependent on A β production but also on A β catabolism. One of the major enzymes being involved in A β degradation is

the insulin-degrading enzyme (IDE) (222). Phospholipids containing DHA (22:6) compared to phospholipids with the corresponding saturated FA (22:0) have been shown to directly bind to IDE (136). In addition purified recombinant IDE enzyme showed an elevated activity in the presence of DHA. Besides its direct effect, DHA enhances exosomal IDE secretion in the extracellular space and altered intracellular/extracellular protein level ratio (136). The effect of DHA on IDE results in an increased A β degradation further enhancing the effect of a decreased A β level in presence of DHA (136). Notably, DHA increases the binding of A β 1-42 to lipid rafts and it has been suggested that DHA might therefore contribute to the clearance of circulating A β by lipid raft dependent degradation pathways (223).

Apart from its impact on A β homeostasis another study revealed a critical role of DHA in the Act-mTOR-S6K pathway in neuronal development and axon outgrowth (224). Moreover, DHA-containing phosphatidyl-choline improved the cognitive deficits in an AD rat model. The authors suggest as a possible mechanism a decreased phosphorylation of Tau in cortex and hippocampal CA1 area (225). *In vitro* studies also indicate that DHA could inhibit and even reverse the formation of A β oligomers and therefore decreases A β -associated neurotoxicity (226-228), which could be recently confirmed in an APP/PS1 rat model of AD emphasizing that dietary DHA modulates A β oligomerisation (229). Moreover a recent study showed that DHA levels were associated with cerebral amyloidosis and preservation of entorhinal hippocampal volumes (215). Notably, the PUFA DHA has been also found to be an agonist of several nuclear receptors, e.g. the retinoic X receptor (RXR) (230, 231) and peroxisome-proliferator receptor (PPAR) α and γ (232-234), a mechanism that might contribute to some of the beneficial effects of DHA in AD. In mouse brain DHA has been identified as endogenous ligand for RXR (230, 231), a ligand-activated nuclear transcription factor, important for reproduction, cellular differentiation, bone development, hematopoiesis and pattern formation during embryogenesis. DHA was found to be specific for RXR, whereas DHA failed to activate the retinoic acid receptor (RAR), the thyroid receptor or the vitamin D receptor (230). These data suggest that DHA influences neuronal function through the activation of a RXR signaling pathway. The ability to activate RXR seems to be not exclusive for DHA, as additional unsaturated fatty acids like docosatetraenoic acid,

docosapentaenoic acid, arachidonic acid and oleic acids have been found to bind and to activate RXR (230, 231). Recently, it has been shown that DHA in combination with bexarotene, a RXR agonist, strongly induces expression of the liver X receptor (LXR):RXR target genes ABCA1 and apoE, involved in reverse cholesterol transport, in a 5XFAD mouse model (235). Furthermore the dual therapy of bexarotene and DHA reduced amyloid pathology, inflammatory processes and restored the impaired working memory of these transgenic mice.

In the paragraph dealing with SM we have already pointed out that a connection between depression and AD, mediated by changes in aSMase or the SM/Cer ratio, exists. Notably n3-FAs and especially EPA and DHA are also associated with major depressive disorder suggesting independently a common lipid-based linkage between depression and AD from another related perspective (236).

It has to be pointed out that dietary DHA supplementation in transgenic AD mouse model did not only increase the DHA level; further other changes in FAs were found, e.g. a reciprocal alteration of DHA and arachidonate was reported (237). Therefore the effect of DHA should not be seen as an isolated event but has to be interpreted in a complex context with other lipid changes (237), also including lipid-based hormonal factors like estrogen also being involved in AD (238). In this context, a recent publication showed that oxidized DHA does not only attenuate the beneficial effect of DHA but even reverses its action leading to increased amyloidogenic pathways (239). 1% oxidized DHA in presence of 99% unoxidized DHA was sufficient to increase A β production in neuroblastoma cells. Interestingly not only one species of oxidized DHA revealed a high amyloidogenic potency, all seven oxidized DHA or lipid peroxidation products elevated the production of A β (239). These results strongly emphasize the need to prevent oxidative reactions of DHA e.g. by adding additional antioxidants. In this context it has to be mentioned that the hydroxylated form of DHA (DHA-H) has been recently reported to improve behavioral motor function and survival of transgenic flies, expressing human A β 42 and tau (240), whereas the ethyl ester form of DHA only showed moderate effects. Orally administration of DHA-H to 5XFAD mice for 4 months also improved cognitive scores of these mice evaluated by the radial arm maze (RAM) test. In human *post mortem* brain an increased level of ROS is a well-known pathological characteristic of AD (241-244). Because of its structure,

DHA is highly susceptible to oxidative stress. Further studies are needed to evaluate if dietary DHA given especially in advanced stages of AD are mainly oxidized and if the beneficial effect of DHA or the controversial effect of oxidized DHA predominates. This also accentuates the question at which stage of disease DHA supplementation might be beneficial.

The important role of DHA during fetal neuronal development is well documented, validating an actual need for DHA early in life (245). An impact of DHA later in life on cognition has been reported (246), but remains controversial especially in respect to AD (247). DHA is required for neuronal function, signaling and neuroprotection in general (248-250). In AD this situation is aggravated because DHA is reduced (251, 252), at least in part, due to reduced supply through liver and diet and a higher need and turnover due to the ongoing neurodegenerative process (253-255). Epidemiological studies highlighted that seafood consumption is associated with slower cognitive decline in apoE4 carriers (256), which is in agreement with the conclusion of a recent meta-analysis that reported that marine derived DHA is associated with a lower risk for AD (257). AD clinical trials turned out negative, subgroup analysis however revealed interesting details. Mild to moderate AD patients who were apoE4 negative, but not apoE4 carriers, as well as patients with very mild AD, showed reduced cognitive decline (258, 259). Nevertheless, the benefit observed was very small; suggesting that overall effectiveness of the DHA is quite modest or even absent in the presence of additional factors that augment the disease process, like apoE4 or an advanced disease phase. This basically leaves two options, to initiate treatment earlier (260) and to enhance the effectiveness of DHA by other means.

Multi-nutrients

Multiple dietary molecules have been suggested to target AD relevant pathologies, e.g. B-vitamins to reduce brain atrophy (261-264), eicosapentaenoic acid (EPA) to boost or exceed the anti-depressive, anti-inflammatory and vascular benefits of DHA (265), phytosterols to counteract the cholesterol effect (112, 115), choline and uridine to improve cognition (266), vitamins E/C and selenium as candidates for AD intervention due to their involvement in oxidative processes (267). Diet *per se* is a multi-nutrient. It therefore is plausible to combine several of those molecules with the aim to achieve a

neuroprotective effect build on the properties of each individual nutrient. That in this context increased effectiveness can indeed be achieved is evidenced e.g. by the combined treatment with choline and uridine which applied together increase synaptic protein levels more than the individual molecules alone (268). A suitable starting point for an AD targeting multi-nutrient diet could be DHA, as the candidate nutrients mentioned above may synergistically work together with DHA. DHA increases spine density, but this can be enhanced by addition of uridine monophosphate (250, 269). Similarly enhancement of cognitive and neuroprotective effects of choline and uridine in combination with DHA (266, 270), or of combined B/E/C-vitamins, selenium, choline, EPA and DHA were noticed (271, 272). In Fortasyn (FC), a multi-nutrient containing all of those nutrients listed above, a complex dietary formulation has been used, which often showed *in vivo* enhanced effects over single nutrients or less complex formulations. In an elegant *in vitro* assay on carbachol-induced membrane potential Savelkoul and colleagues showed that for treatment with either of the above-mentioned vitamins, selenium, choline, uridine and phospholipids none showed any response. Only DHA and EPA did. Nonetheless, when these molecules were successively added to DHA the membrane potential change increased (273), providing an example for the important role of DHA to activate the synergistic potential of those dietary molecules and that the effectiveness of specific nutrients may depend on the dietary context in which they are provided (274).

A linear model has recently been proposed to explain this observation (275, 276). In this model these molecules serve as precursors and co-factors for DHA-enriched membrane synthesis. This would then result in synapse fortification and eventually memory improvement, combining several biochemical and *in vivo* findings into a more complex model (275). In addition to possibly working through a single cascading biochemical pathway, multiple nutrients will target a number of different cellular processes, some of which may be disease relevant. Koivisto recently compared fish oil (predominantly DHA and EPA), fish oil supplemented with the plant sterol stigmasterol or FC in a head-to-head study in 14 month old APP/PS transgenic mice (115). Neither diet affected the APP/PS induced hyperactivity, but all diets improved odor recognition. Only FC had an effect on spatial learning, while only stigmasterol improved habituation and suppressed microglial activation. In another approach treatment with DHA+EPA+uridine largely, and with FC almost entirely, ameliorated white and gray matter structural

integrity. FC was more effective on aspects of neuronal repair and maintenance, neurogenesis and anxiety-related behavior. Other AD related pathologies targeted by this multi-nutrient approach include cerebral blood flow and perfusion, secretase activity, amyloid pathology, hippocampal neurotransmission and apoE4 related pathologies (36, 115, 277-281). Taken together these results indicate that lipid based multi-target based approaches have the potential to be more effective than treatment with single lipids. While dietary habits have been extensively studied in AD and concluded that healthy dietary patterns, e.g. adherence to a Mediterranean diet or regular fish consumption may reduce the dementia risk (257, 282), few clinical trials studied specific multi-nutrient intervention. B-vitamin supplementation in patients with mild cognitive impairment in a post-hoc analysis resulted in reduced brain atrophy and increased cognitive improvement, which was limited to that subgroup of patients which had the highest blood-level of n3-FAs, especially DHA (264, 283, 284). A set of four clinical trials based on the FC multi-nutrient formulation, showed improved memory performance (285, 286), increased neurophysiological measures of synaptic activity, and enhanced functional connectivity in the brain (286, 287), but did not slow cognitive decline in mild to moderate AD (288, 289). Results from the fourth clinical trial, which studies this intervention in prodromal (pre dementia) AD have not been published thus far.

It is now evident that lipids play an important role in AD. Presence of the apoE4 allele may provide serious challenges as well as on the way towards Successful implementation in therapeutic approaches will require improving effectiveness over what has been achieved currently. Multi-nutrients, earlier intervention within the AD continuum and combination with other pharmaceutical and non-pharmaceutica interventions may provide future steps in this direction.

Acknowledgments/grant support:

The research leading to these results has received funding from the EU FP7 project LipiDiDiet, Grant Agreement No. 211696. Moreover funding for MOWG and TH was provided by Fundació la Marató de TV3 and by JPND MindAD 1ED1508.

References:

1. Maresova, P., H. Mohelska, J. Dolejs, and K. Kuca. 2015. Socio-economic Aspects of Alzheimer's Disease. *Curr Alzheimer Res* **12**: 903-911.
2. Mattson, M. P. 2004. Pathways towards and away from Alzheimer's disease. *Nature* **430**: 631-639.
3. Foley, P. 2010. Lipids in Alzheimer's disease: A century-old story. *Biochim Biophys Acta* **1801**: 750-753.
4. Alzheimer, A. 1907. Über eine eigenartige Erkrankung der Hirnrinde. *Allg. Z. Psychiatrie Psychisch-Gerichtlich. Med.* **64**: 146-148.
5. Corder, E. H., A. M. Saunders, W. J. Strittmatter, D. E. Schmechel, P. C. Gaskell, G. W. Small, A. D. Roses, J. L. Haines, and M. A. Pericak-Vance. 1993. Gene dose of apolipoprotein E type 4 allele and the risk of Alzheimer's disease in late onset families. *Science* **261**: 921-923.
6. Czech, C., V. Monning, P. J. Tienari, T. Hartmann, C. Masters, K. Beyreuther, and H. Forstl. 1993. Apolipoprotein E-epsilon 4 allele and Alzheimer's disease. *Lancet* **342**: 1309.
7. Ueki, A., M. Kawano, Y. Namba, M. Kawakami, and K. Ikeda. 1993. A high frequency of apolipoprotein E4 isoprotein in Japanese patients with late-onset nonfamilial Alzheimer's disease. *Neurosci Lett* **163**: 166-168.
8. Mayeux, R., R. Ottman, M. X. Tang, L. Noboa-Bauza, K. Marder, B. Gurland, and Y. Stern. 1993. Genetic susceptibility and head injury as risk factors for Alzheimer's disease among community-dwelling elderly persons and their first-degree relatives. *Ann Neurol* **33**: 494-501.
9. Sparks, D. L., S. W. Scheff, J. C. Hunsaker, 3rd, H. Liu, T. Landers, and D. R. Gross. 1994. Induction of Alzheimer-like beta-amyloid immunoreactivity in the brains of rabbits with dietary cholesterol. *Exp Neurol* **126**: 88-94.

10. Refolo, L. M., M. A. Pappolla, B. Malester, J. LaFrancois, T. Bryant-Thomas, R. Wang, G. S. Tint, K. Sambamurti, and K. Duff. 2000. Hypercholesterolemia accelerates the Alzheimer's amyloid pathology in a transgenic mouse model. *Neurobiol Dis* **7**: 321-331.
11. Shie, F. S., L. W. Jin, D. G. Cook, J. B. Leverenz, and R. C. LeBoeuf. 2002. Diet-induced hypercholesterolemia enhances brain A beta accumulation in transgenic mice. *Neuroreport* **13**: 455-459.
12. Simons, M., B. de Strooper, G. Multhaup, P. J. Tienari, C. G. Dotti, and K. Beyreuther. 1996. Amyloidogenic processing of the human amyloid precursor protein in primary cultures of rat hippocampal neurons. *J-Neurosci* **16**: 899-908.
13. Simons, M., P. Keller, B. De Strooper, K. Beyreuther, C. G. Dotti, and K. Simons. 1998. Cholesterol depletion inhibits the generation of beta-amyloid in hippocampal neurons. *Proc Natl Acad Sci U S A* **95**: 6460-6464.
14. Fassbender, K., M. Simons, C. Bergmann, M. Stroick, D. Lutjohann, P. Keller, H. Runz, S. Kuhl, T. Bertsch, K. von Bergmann, M. Hennerici, K. Beyreuther, and T. Hartmann. 2001. Simvastatin strongly reduces levels of Alzheimer's disease beta -amyloid peptides Abeta 42 and Abeta 40 in vitro and in vivo. *Proc Natl Acad Sci U S A* **98**: 5856-5861.
15. Wolozin, B., W. Kellman, P. Ruosseau, G. G. Celesia, and G. Siegel. 2000. Decreased prevalence of alzheimer disease associated with 3-hydroxy-3- methylglutaryl coenzyme A reductase inhibitors. *Arch Neurol* **57**: 1439-1443.
16. Kivipelto, M., E. L. Helkala, M. P. Laakso, T. Hanninen, M. Hallikainen, K. Alhainen, H. Soininen, J. Tuomilehto, and A. Nissinen. 2001. Midlife vascular risk factors and Alzheimer's disease in later life: longitudinal, population based study. *Bmj* **322**: 1447-1451.
17. Sano, M., K. L. Bell, D. Galasko, J. E. Galvin, R. G. Thomas, C. H. van Dyck, and P. S. Aisen. 2011. A randomized, double-blind, placebo-controlled trial of simvastatin to treat Alzheimer disease. *Neurology* **77**: 556-563.

18. Feldman, H. H., R. S. Doody, M. Kivipelto, D. L. Sparks, D. D. Waters, R. W. Jones, E. Schwam, R. Schindler, J. Hey-Hadavi, D. A. DeMicco, and A. Breazna. 2010. Randomized controlled trial of atorvastatin in mild to moderate Alzheimer disease: LEADe. *Neurology* **74**: 956-964.
19. Saunders, A. M., W. J. Strittmatter, D. Schmechel, P. H. George-Hyslop, M. A. Pericak-Vance, S. H. Joo, B. L. Rosi, J. F. Gusella, D. R. Crapper-MacLachlan, M. J. Alberts, and et al. 1993. Association of apolipoprotein E allele epsilon 4 with late-onset familial and sporadic Alzheimer's disease. *Neurology* **43**: 1467-1472.
20. Roses, A. D. 1996. Apolipoprotein E alleles as risk factors in Alzheimer's disease. *Annual review of medicine* **47**: 387-400.
21. Harold, D., R. Abraham, P. Hollingworth, R. Sims, A. Gerrish, M. L. Hamshere, J. S. Pahwa, V. Moskvina, K. Dowzell, A. Williams, N. Jones, C. Thomas, A. Stretton, A. R. Morgan, S. Lovestone, J. Powell, P. Proitsi, M. K. Lupton, C. Brayne, D. C. Rubinsztein, M. Gill, B. Lawlor, A. Lynch, K. Morgan, K. S. Brown, P. A. Passmore, D. Craig, B. McGuinness, S. Todd, C. Holmes, D. Mann, A. D. Smith, S. Love, P. G. Kehoe, J. Hardy, S. Mead, N. Fox, M. Rossor, J. Collinge, W. Maier, F. Jessen, B. Schurmann, R. Heun, H. van den Bussche, I. Heuser, J. Kornhuber, J. Wiltfang, M. Dichgans, L. Frolich, H. Hampel, M. Hull, D. Rujescu, A. M. Goate, J. S. Kauwe, C. Cruchaga, P. Nowotny, J. C. Morris, K. Mayo, K. Sleegers, K. Bettens, S. Engelborghs, P. P. De Deyn, C. Van Broeckhoven, G. Livingston, N. J. Bass, H. Gurling, A. McQuillin, R. Gwilliam, P. Deloukas, A. Al-Chalabi, C. E. Shaw, M. Tsolaki, A. B. Singleton, R. Guerreiro, T. W. Muhleisen, M. M. Nothen, S. Moebus, K. H. Jockel, N. Klopp, H. E. Wichmann, M. M. Carrasquillo, V. S. Pankratz, S. G. Younkin, P. A. Holmans, M. O'Donovan, M. J. Owen, and J. Williams. 2009. Genome-wide association study identifies variants at CLU and PICALM associated with Alzheimer's disease. *Nat Genet* **41**: 1088-1093.
22. Jones, L., P. A. Holmans, M. L. Hamshere, D. Harold, V. Moskvina, D. Ivanov, A. Pocklington, R. Abraham, P. Hollingworth, R. Sims, A. Gerrish, J. S. Pahwa, N. Jones, A. Stretton, A. R. Morgan, S. Lovestone, J. Powell, P. Proitsi, M. K. Lupton, C. Brayne, D. C. Rubinsztein, M. Gill, B. Lawlor, A. Lynch, K. Morgan, K. S. Brown, P. A. Passmore, D. Craig, B. McGuinness, S. Todd, C. Holmes, D. Mann, A. D. Smith, S. Love, P. G. Kehoe, S. Mead, N. Fox, M. Rossor, J. Collinge, W. Maier, F. Jessen, B.

- Schurmann, R. Heun, H. Kolsch, H. van den Bussche, I. Heuser, O. Peters, J. Kornhuber, J. Wiltfang, M. Dichgans, L. Frolich, H. Hampel, M. Hull, D. Rujescu, A. M. Goate, J. S. Kauwe, C. Cruchaga, P. Nowotny, J. C. Morris, K. Mayo, G. Livingston, N. J. Bass, H. Gurling, A. McQuillin, R. Gwilliam, P. Deloukas, A. Al-Chalabi, C. E. Shaw, A. B. Singleton, R. Guerreiro, T. W. Muhleisen, M. M. Nothen, S. Moebus, K. H. Jockel, N. Klopp, H. E. Wichmann, E. Ruther, M. M. Carrasquillo, V. S. Pankratz, S. G. Younkin, J. Hardy, M. C. O'Donovan, M. J. Owen, and J. Williams. 2010. Genetic evidence implicates the immune system and cholesterol metabolism in the aetiology of Alzheimer's disease. *PLoS one* **5**: e13950.
23. Ghiso, J., E. Matsubara, A. Koudinov, N. H. Choi-Miura, M. Tomita, T. Wisniewski, and B. Frangione. 1993. The cerebrospinal-fluid soluble form of Alzheimer's amyloid beta is complexed to SP-40,40 (apolipoprotein J), an inhibitor of the complement membrane-attack complex. *Biochem J* **293 (Pt 1)**: 27-30.
24. Zlokovic, B. V., C. L. Martel, J. B. Mackic, E. Matsubara, T. Wisniewski, J. G. McComb, B. Frangione, and J. Ghiso. 1994. Brain uptake of circulating apolipoproteins J and E complexed to Alzheimer's amyloid beta. *Biochem Biophys Res Commun* **205**: 1431-1437.
25. Dallongeville, J., S. Lussier-Cacan, and J. Davignon. 1992. Modulation of plasma triglyceride levels by apoE phenotype: a meta-analysis. *Journal of lipid research* **33**: 447-454.
26. Hospattankar, A. V., T. Fairwell, R. Ronan, and H. B. Brewer, Jr. 1984. Amino acid sequence of human plasma apolipoprotein C-II from normal and hyperlipoproteinemic subjects. *The Journal of biological chemistry* **259**: 318-322.
27. Hanson, A. J., J. L. Bayer-Carter, P. S. Green, T. J. Montine, C. W. Wilkinson, L. D. Baker, G. S. Watson, L. M. Bonner, M. Callaghan, J. B. Leverenz, E. Tsai, N. Postupna, J. Zhang, J. Lampe, and S. Craft. 2013. Effect of apolipoprotein E genotype and diet on apolipoprotein E lipidation and amyloid peptides: randomized clinical trial. *JAMA neurology* **70**: 972-980.
28. Notkola, I. L., R. Sulkava, J. Pekkanen, T. Erkinjuntti, C. Ehnholm, P. Kivinen, J. Tuomilehto, and A. Nissinen. 1998. Serum total cholesterol, apolipoprotein E epsilon 4 allele, and Alzheimer's disease. *Neuroepidemiology* **17**: 14-20.

29. Pappolla, M. A., T. K. Bryant-Thomas, D. Herbert, J. Pacheco, M. Fabra Garcia, M. Manjon, X. Girones, T. L. Henry, E. Matsubara, D. Zambon, B. Woloizin, M. Sano, F. F. Cruz-Sanchez, L. J. Thal, S. S. Petanceska, and L. M. Refolo. 2003. Mild hypercholesterolemia is an early risk factor for the development of Alzheimer amyloid pathology. *Neurology* **61**: 199-205.
30. Kivipelto, M., E. L. Helkala, M. P. Laakso, T. Hanninen, M. Hallikainen, K. Alhainen, S. Iivonen, A. Mannermaa, J. Tuomilehto, A. Nissinen, and H. Soininen. 2002. Apolipoprotein E epsilon4 allele, elevated midlife total cholesterol level, and high midlife systolic blood pressure are independent risk factors for late-life Alzheimer disease. *Ann Intern Med* **137**: 149-155.
31. Mulder, M., R. Ravid, D. F. Swaab, E. R. de Kloet, E. D. Haasdijk, J. Julk, J. J. van der Boom, and L. M. Havekes. 1998. Reduced levels of cholesterol, phospholipids, and fatty acids in cerebrospinal fluid of Alzheimer disease patients are not related to apolipoprotein E4. *Alzheimer disease and associated disorders* **12**: 198-203.
32. Couttas, T. A., N. Kain, B. Daniels, X. Y. Lim, C. Shepherd, J. Kril, R. Pickford, H. Li, B. Garner, and A. S. Don. 2014. Loss of the neuroprotective factor Sphingosine 1-phosphate early in Alzheimer's disease pathogenesis. *Acta Neuropathol Commun* **2**: 9.
33. Zhu, L., M. Zhong, G. A. Elder, M. Sano, D. M. Holtzman, S. Gandy, C. Cardozo, V. Haroutunian, N. K. Robakis, and D. Cai. 2015. Phospholipid dysregulation contributes to ApoE4-associated cognitive deficits in Alzheimer's disease pathogenesis. *Proc Natl Acad Sci U S A* **112**: 11965-11970.
34. Kariv-Inbal, Z., S. Yacobson, R. Berkecz, M. Peter, T. Janaky, D. Lutjohann, L. M. Broersen, T. Hartmann, and D. M. Michaelson. 2012. The isoform-specific pathological effects of apoE4 in vivo are prevented by a fish oil (DHA) diet and are modified by cholesterol. *J Alzheimers Dis* **28**: 667-683.
35. Sharman, M. J., G. Shui, A. Z. Fernandis, W. L. Lim, T. Berger, E. Hone, K. Taddei, I. J. Martins, J. Ghiso, J. D. Buxbaum, S. Gandy, M. R. Wenk, and R. N. Martins. 2010. Profiling brain and plasma lipids in human APOE epsilon2, epsilon3, and epsilon4 knock-in mice using electrospray ionization mass spectrometry. *J Alzheimers Dis* **20**: 105-111.

36. Wiesmann, M., V. Zerbi, D. Jansen, R. Haast, D. Lutjohann, L. M. Broersen, A. Heerschap, and A. J. Kiliaan. 2016. A Dietary Treatment Improves Cerebral Blood Flow and Brain Connectivity in Aging apoE4 Mice. *Neural plasticity* **2016**: 6846721.
37. Maioli, S., E. Puerta, P. Merino-Serrais, L. Fusari, F. Gil-Bea, R. Rimondini, and A. Cedazo-Minguez. 2012. Combination of apolipoprotein E4 and high carbohydrate diet reduces hippocampal BDNF and arc levels and impairs memory in young mice. *J Alzheimers Dis* **32**: 341-355.
38. Weisgraber, K. H. 1994. Apolipoprotein E: structure-function relationships. *Adv Protein Chem* **45**: 249-302.
39. Samieri, C., S. Lorrain, B. Buaud, C. Vaysse, C. Berr, E. Peuchant, S. C. Cunnane, and P. Barberger-Gateau. 2013. Relationship between diet and plasma long-chain n-3 PUFAs in older people: impact of apolipoprotein E genotype. *Journal of lipid research* **54**: 2559-2567.
40. Heinsinger, N. M., M. A. Gachechiladze, and G. W. Rebeck. 2016. Apolipoprotein E Genotype Affects Size of ApoE Complexes in Cerebrospinal Fluid. *Journal of neuropathology and experimental neurology* **75**: 918-924.
41. Hu, J., C. C. Liu, X. F. Chen, Y. W. Zhang, H. Xu, and G. Bu. 2015. Opposing effects of viral mediated brain expression of apolipoprotein E2 (apoE2) and apoE4 on apoE lipidation and Abeta metabolism in apoE4-targeted replacement mice. *Molecular neurodegeneration* **10**: 6.
42. Oram, J. F., and A. M. Vaughan. 2006. ATP-Binding cassette cholesterol transporters and cardiovascular disease. *Circulation research* **99**: 1031-1043.
43. Krimbou, L., M. Denis, B. Haidar, M. Carrier, M. Marcil, and J. Genest, Jr. 2004. Molecular interactions between apoE and ABCA1: impact on apoE lipidation. *Journal of lipid research* **45**: 839-848.
44. Kennedy, M. A., G. C. Barrera, K. Nakamura, A. Baldan, P. Tarr, M. C. Fishbein, J. Frank, O. L. Francone, and P. A. Edwards. 2005. ABCG1 has a critical role in mediating cholesterol efflux to HDL and preventing cellular lipid accumulation. *Cell metabolism* **1**: 121-131.

45. Nordestgaard, L. T., A. Tybjaerg-Hansen, B. G. Nordestgaard, and R. Frikke-Schmidt. 2015. Loss-of-function mutation in ABCA1 and risk of Alzheimer's disease and cerebrovascular disease. *Alzheimer's & dementia : the journal of the Alzheimer's Association* **11**: 1430-1438.
46. Wahrle, S. E., H. Jiang, M. Parsadanian, J. Legleiter, X. Han, J. D. Fryer, T. Kowalewski, and D. M. Holtzman. 2004. ABCA1 is required for normal central nervous system ApoE levels and for lipidation of astrocyte-secreted apoE. *The Journal of biological chemistry* **279**: 40987-40993.
47. Swift, L. L., M. H. Farkas, A. S. Major, K. Valyi-Nagy, M. F. Linton, and S. Fazio. 2001. A recycling pathway for resecretion of internalized apolipoprotein E in liver cells. *The Journal of biological chemistry* **276**: 22965-22970.
48. Hirsch-Reinshagen, V., S. Zhou, B. L. Burgess, L. Bernier, S. A. McIsaac, J. Y. Chan, G. H. Tansley, J. S. Cohn, M. R. Hayden, and C. L. Wellington. 2004. Deficiency of ABCA1 impairs apolipoprotein E metabolism in brain. *The Journal of biological chemistry* **279**: 41197-41207.
49. Boehm-Cagan, A., and D. M. Michaelson. 2014. Reversal of apoE4-driven brain pathology and behavioral deficits by bexarotene. *The Journal of neuroscience : the official journal of the Society for Neuroscience* **34**: 7293-7301.
50. Boehm-Cagan, A., R. Bar, O. Liraz, J. K. Bielicki, J. O. Johansson, and D. M. Michaelson. 2016. ABCA1 Agonist Reverses the ApoE4-Driven Cognitive and Brain Pathologies. *J Alzheimers Dis* **54**: 1219-1233.
51. Hafiane, A., J. K. Bielicki, J. O. Johansson, and J. Genest. 2015. Novel Apo E-Derived ABCA1 Agonist Peptide (CS-6253) Promotes Reverse Cholesterol Transport and Induces Formation of prebeta-1 HDL In Vitro. *PloS one* **10**: e0131997.
52. Wahrle, S. E., H. Jiang, M. Parsadanian, J. Kim, A. Li, A. Knoten, S. Jain, V. Hirsch-Reinshagen, C. L. Wellington, K. R. Bales, S. M. Paul, and D. M. Holtzman. 2008. Overexpression of ABCA1 reduces amyloid deposition in the PDAPP mouse model of Alzheimer disease. *J Clin Invest* **118**: 671-682.
53. Nakato, M., M. Matsuo, N. Kono, M. Arita, H. Arai, J. Ogawa, N. Kioka, and K. Ueda. 2015. Neurite outgrowth stimulation by n-3 and n-6 PUFAs of phospholipids in apoE-containing lipoproteins secreted from glial cells. *J Lipid Res* **56**: 1880-1890.

54. LaDu, M. J., M. T. Falduto, A. M. Manelli, C. A. Reardon, G. S. Getz, and D. E. Frail. 1994. Isoform-specific binding of apolipoprotein E to beta-amyloid. *The Journal of biological chemistry* **269**: 23403-23406.
55. Tokuda, T., M. Calero, E. Matsubara, R. Vidal, A. Kumar, B. Permanne, B. Zlokovic, J. D. Smith, M. J. Ladu, A. Rostagno, B. Frangione, and J. Ghiso. 2000. Lipidation of apolipoprotein E influences its isoform-specific interaction with Alzheimer's amyloid beta peptides. *The Biochemical journal* **348 Pt 2**: 359-365.
56. Kim, J., J. M. Basak, and D. M. Holtzman. 2009. The role of apolipoprotein E in Alzheimer's disease. *Neuron* **63**: 287-303.
57. Verghese, P. B., J. M. Castellano, K. Garai, Y. Wang, H. Jiang, A. Shah, G. Bu, C. Frieden, and D. M. Holtzman. 2013. ApoE influences amyloid-beta (A β) clearance despite minimal apoE/A β association in physiological conditions. *Proc Natl Acad Sci U S A* **110**: E1807-1816.
58. Holtzman, D. M., J. Herz, and G. Bu. 2012. Apolipoprotein E and apolipoprotein E receptors: normal biology and roles in Alzheimer disease. *Cold Spring Harbor perspectives in medicine* **2**: a006312.
59. Liraz, O., A. Boehm-Cagan, and D. M. Michaelson. 2013. ApoE4 induces A β 42, tau, and neuronal pathology in the hippocampus of young targeted replacement apoE4 mice. *Molecular neurodegeneration* **8**: 16.
60. Mahley, R. W., and S. C. Rall, Jr. 2000. Apolipoprotein E: far more than a lipid transport protein. *Annual review of genomics and human genetics* **1**: 507-537.
61. Nathan, B. P., Y. Jiang, G. K. Wong, F. Shen, G. J. Brewer, and R. G. Struble. 2002. Apolipoprotein E4 inhibits, and apolipoprotein E3 promotes neurite outgrowth in cultured adult mouse cortical neurons through the low-density lipoprotein receptor-related protein. *Brain research* **928**: 96-105.
62. Nathan, B. P., S. Bellosta, D. A. Sanan, K. H. Weisgraber, R. W. Mahley, and R. E. Pitas. 1994. Differential effects of apolipoproteins E3 and E4 on neuronal growth in vitro. *Science* **264**: 850-852.

63. Bellosta, S., B. P. Nathan, M. Orth, L. M. Dong, R. W. Mahley, and R. E. Pitas. 1995. Stable expression and secretion of apolipoproteins E3 and E4 in mouse neuroblastoma cells produces differential effects on neurite outgrowth. *The Journal of biological chemistry* **270**: 27063-27071.
64. Michaelson, D. M. 2014. APOE epsilon4: the most prevalent yet understudied risk factor for Alzheimer's disease. *Alzheimer's & dementia : the journal of the Alzheimer's Association* **10**: 861-868.
65. Glenner, G. G., and C. W. Wong. 2012. Alzheimer's disease: initial report of the purification and characterization of a novel cerebrovascular amyloid protein. 1984. *Biochem Biophys Res Commun* **425**: 534-539.
66. Masters, C. L., G. Simms, N. A. Weinman, G. Multhaup, B. L. McDonald, and K. Beyreuther. 1985. Amyloid plaque core protein in Alzheimer disease and Down syndrome. *Proc Natl Acad Sci U S A* **82**: 4245-4249.
67. Grundke-Iqbal, I., K. Iqbal, M. Quinlan, Y. C. Tung, M. S. Zaidi, and H. M. Wisniewski. 1986. Microtubule-associated protein tau. A component of Alzheimer paired helical filaments. *J Biol Chem* **261**: 6084-6089.
68. Grundke-Iqbal, I., K. Iqbal, Y. C. Tung, M. Quinlan, H. M. Wisniewski, and L. I. Binder. 1986. Abnormal phosphorylation of the microtubule-associated protein tau (tau) in Alzheimer cytoskeletal pathology. *Proc Natl Acad Sci U S A* **83**: 4913-4917.
69. Dyrks, T., A. Weidemann, G. Multhaup, J. M. Salbaum, H. G. Lemaire, J. Kang, B. Muller-Hill, C. L. Masters, and K. Beyreuther. 1988. Identification, transmembrane orientation and biogenesis of the amyloid A4 precursor of Alzheimer's disease. *EMBO J* **7**: 949-957.
70. Sinha, S., J. P. Anderson, R. Barbour, G. S. Basi, R. Caccavello, D. Davis, M. Doan, H. F. Dovey, N. Frigon, J. Hong, K. Jacobson-Croak, N. Jewett, P. Keim, J. Knops, I. Lieberburg, M. Power, H. Tan, G. Tatsuno, J. Tung, D. Schenk, P. Seubert, S. M. Suomensaaari, S. Wang, D. Walker, J. Zhao, L. McConlogue, and V. John. 1999. Purification and cloning of amyloid precursor protein beta-secretase from human brain. *Nature* **402**: 537-540.
71. Vassar, R., B. D. Bennett, S. Babu-Khan, S. Kahn, E. A. Mendiaz, P. Denis, D. B. Teplow, S. Ross, P. Amarante, R. Loeloff, Y. Luo, S. Fisher, J. Fuller, S. Edenson, J. Lile, M. A. Jarosinski, A. L. Biere, E.

Curran, T. Burgess, J. C. Louis, F. Collins, J. Treanor, G. Rogers, and M. Citron. 1999. Beta-secretase cleavage of Alzheimer's amyloid precursor protein by the transmembrane aspartic protease BACE. *Science* **286**: 735-741.

72. Haass, C. 2004. Take five--BACE and the gamma-secretase quartet conduct Alzheimer's amyloid beta-peptide generation. *EMBO J* **23**: 483-488.

73. Haass, C., M. G. Schlossmacher, A. Y. Hung, C. Vigo-Pelfrey, A. Mellon, B. L. Ostaszewski, I. Lieberburg, E. H. Koo, D. Schenk, D. B. Teplow, and et al. 1992. Amyloid beta-peptide is produced by cultured cells during normal metabolism. *Nature* **359**: 322-325.

74. von Rotz, R. C., B. M. Kohli, J. Bosset, M. Meier, T. Suzuki, R. M. Nitsch, and U. Konietzko. 2004. The APP intracellular domain forms nuclear multiprotein complexes and regulates the transcription of its own precursor. *J Cell Sci* **117**: 4435-4448.

75. Cao, X., and T. C. Sudhof. 2001. A transcriptionally [correction of transcriptively] active complex of APP with Fe65 and histone acetyltransferase Tip60. *Science* **293**: 115-120.

76. Grimm, M. O., S. Grosgen, T. L. Rothhaar, V. K. Burg, B. Hundsdorfer, V. J. Haupenthal, P. Friess, U. Muller, K. Fassbender, M. Riemenschneider, H. S. Grimm, and T. Hartmann. 2011. Intracellular APP Domain Regulates Serine-Palmitoyl-CoA Transferase Expression and Is Affected in Alzheimer's Disease. *Int J Alzheimers Dis* **2011**: 695413.

77. Grimm, M. O., J. Kuchenbecker, T. L. Rothhaar, S. Grosgen, B. Hundsdorfer, V. K. Burg, P. Friess, U. Muller, H. S. Grimm, M. Riemenschneider, and T. Hartmann. 2011. Plasmalogen synthesis is regulated via alkyl-dihydroxyacetonephosphate-synthase by amyloid precursor protein processing and is affected in Alzheimer's disease. *J Neurochem* **116**: 916-925.

78. Grimm, M. O., E. G. Zinser, S. Grosgen, B. Hundsdorfer, T. L. Rothhaar, V. K. Burg, L. Kaestner, T. A. Bayer, P. Lipp, U. Muller, H. S. Grimm, and T. Hartmann. 2012. Amyloid precursor protein (APP) mediated regulation of ganglioside homeostasis linking Alzheimer's disease pathology with ganglioside metabolism. *PLoS One* **7**: e34095.

79. Robinson, A., S. Grosgen, J. Mett, V. C. Zimmer, V. J. Haupenthal, B. Hundsdorfer, C. P. Stahlmann, Y. Slobodskoy, U. C. Muller, T. Hartmann, R. Stein, and M. O. Grimm. 2014. Upregulation

of PGC-1 α expression by Alzheimer's disease-associated pathway: presenilin 1/amyloid precursor protein (APP)/intracellular domain of APP. *Aging Cell* **13**: 263-272.

80. Grimm, M. O., J. Mett, C. P. Stahlmann, S. Grosgen, V. J. Haupenthal, T. Blumel, B. Hundsdorfer, V. C. Zimmer, N. T. Mylonas, H. Tanila, U. Muller, H. S. Grimm, and T. Hartmann. 2015. APP intracellular domain derived from amyloidogenic beta- and gamma-secretase cleavage regulates neprilysin expression. *Front Aging Neurosci* **7**: 77.

81. Grimm, M. O., J. Mett, C. P. Stahlmann, V. J. Haupenthal, V. C. Zimmer, and T. Hartmann. 2013. Neprilysin and A β Clearance: Impact of the APP Intracellular Domain in NEP Regulation and Implications in Alzheimer's Disease. *Front Aging Neurosci* **5**: 98.

82. Grimm, M. O., B. Hundsdorfer, S. Grosgen, J. Mett, V. C. Zimmer, C. P. Stahlmann, V. J. Haupenthal, T. L. Rothhaar, J. Lehmann, A. Patzold, E. G. Zinser, H. Tanila, J. Shen, U. Muller, H. S. Grimm, and T. Hartmann. 2014. PS dependent APP cleavage regulates glucosylceramide synthase and is affected in Alzheimer's disease. *Cell Physiol Biochem* **34**: 92-110.

83. Pardossi-Piquard, R., and F. Checler. 2012. The physiology of the beta-amyloid precursor protein intracellular domain AICD. *J Neurochem* **120 Suppl 1**: 109-124.

84. Iwatsubo, T., A. Odaka, N. Suzuki, H. Mizusawa, N. Nukina, and Y. Ihara. 1994. Visualization of A β 42(43) and A β 40 in senile plaques with end-specific A β monoclonals: evidence that an initially deposited species is A β 42(43). *Neuron* **13**: 45-53.

85. Seubert, P., C. Vigo-Pelfrey, F. Esch, M. Lee, H. Dovey, D. Davis, S. Sinha, M. Schlossmacher, J. Whaley, C. Swindlehurst, and et al. 1992. Isolation and quantification of soluble Alzheimer's beta-peptide from biological fluids. *Nature* **359**: 325-327.

86. Wang, R., D. Sweeney, S. E. Gandy, and S. S. Sisodia. 1996. The profile of soluble amyloid beta protein in cultured cell media. Detection and quantification of amyloid beta protein and variants by immunoprecipitation-mass spectrometry. *J Biol Chem* **271**: 31894-31902.

87. Schieb, H., H. Kratzin, O. Jahn, W. Mobius, S. Rabe, M. Staufenbiel, J. Wiltfang, and H. W. Klafki. 2011. Beta-amyloid peptide variants in brains and cerebrospinal fluid from amyloid precursor

protein (APP) transgenic mice: comparison with human Alzheimer amyloid. *J Biol Chem* **286**: 33747-33758.

88. Jarrett, J. T., E. P. Berger, and P. T. Lansbury, Jr. 1993. The carboxy terminus of the beta amyloid protein is critical for the seeding of amyloid formation: implications for the pathogenesis of Alzheimer's disease. *Biochemistry* **32**: 4693-4697.

89. Jarrett, J. T., E. P. Berger, and P. T. Lansbury, Jr. 1993. The C-terminus of the beta protein is critical in amyloidogenesis. *Ann N Y Acad Sci* **695**: 144-148.

90. Lesne, S., M. T. Koh, L. Kotilinek, R. Kaye, C. G. Glabe, A. Yang, M. Gallagher, and K. H. Ashe. 2006. A specific amyloid-beta protein assembly in the brain impairs memory. *Nature* **440**: 352-357.

91. Kaye, R., E. Head, J. L. Thompson, T. M. McIntire, S. C. Milton, C. W. Cotman, and C. G. Glabe. 2003. Common structure of soluble amyloid oligomers implies common mechanism of pathogenesis. *Science* **300**: 486-489.

92. Shankar, G. M., S. Li, T. H. Mehta, A. Garcia-Munoz, N. E. Shepardson, I. Smith, F. M. Brett, M. A. Farrell, M. J. Rowan, C. A. Lemere, C. M. Regan, D. M. Walsh, B. L. Sabatini, and D. J. Selkoe. 2008. Amyloid-beta protein dimers isolated directly from Alzheimer's brains impair synaptic plasticity and memory. *Nat Med* **14**: 837-842.

93. Umeda, T., T. Tomiyama, N. Sakama, S. Tanaka, M. P. Lambert, W. L. Klein, and H. Mori. 2011. Intraneuronal amyloid beta oligomers cause cell death via endoplasmic reticulum stress, endosomal/lysosomal leakage, and mitochondrial dysfunction in vivo. *J Neurosci Res* **89**: 1031-1042.

94. Kimberly, W. T., M. J. LaVoie, B. L. Ostaszewski, W. Ye, M. S. Wolfe, and D. J. Selkoe. 2003. Gamma-secretase is a membrane protein complex comprised of presenilin, nicastrin, Aph-1, and Pen-2. *Proc Natl Acad Sci U S A* **100**: 6382-6387.

95. Herreman, A., L. Serneels, W. Annaert, D. Collen, L. Schoonjans, and B. De Strooper. 2000. Total inactivation of gamma-secretase activity in presenilin-deficient embryonic stem cells. *Nat Cell Biol* **2**: 461-462.

96. Nyabi, O., S. Pype, M. Mercken, A. Herreman, P. Saftig, K. Craessaerts, L. Serneels, W. Annaert, and B. De Strooper. 2002. No endogenous A beta production in presenilin-deficient fibroblasts. *Nat Cell Biol* **4**: E164; author reply E165-166.
97. Walker, E. S., M. Martinez, A. L. Brunkan, and A. Goate. 2005. Presenilin 2 familial Alzheimer's disease mutations result in partial loss of function and dramatic changes in Abeta 42/40 ratios. *J Neurochem* **92**: 294-301.
98. Duering, M., M. O. Grimm, H. S. Grimm, J. Schroder, and T. Hartmann. 2005. Mean age of onset in familial Alzheimer's disease is determined by amyloid beta 42. *Neurobiol Aging* **26**: 785-788.
99. Bentahir, M., O. Nyabi, J. Verhamme, A. Tolia, K. Horre, J. Wiltfang, H. Esselmann, and B. De Strooper. 2006. Presenilin clinical mutations can affect gamma-secretase activity by different mechanisms. *J Neurochem* **96**: 732-742.
100. Wiley, J. C., M. Hudson, K. C. Kanning, L. C. Schecterson, and M. Bothwell. 2005. Familial Alzheimer's disease mutations inhibit gamma-secretase-mediated liberation of beta-amyloid precursor protein carboxy-terminal fragment. *J Neurochem* **94**: 1189-1201.
101. Lee, S. J., U. Liyanage, P. E. Bickel, W. Xia, P. T. Lansbury, Jr., and K. S. Kosik. 1998. A detergent-insoluble membrane compartment contains A beta in vivo. *Nat Med* **4**: 730-734.
102. Riddell, D. R., G. Christie, I. Hussain, and C. Dingwall. 2001. Compartmentalization of beta-secretase (Asp2) into low-buoyant density, noncaveolar lipid rafts. *Curr Biol* **11**: 1288-1293.
103. Ehehalt, R., P. Keller, C. Haass, C. Thiele, and K. Simons. 2003. Amyloidogenic processing of the Alzheimer beta-amyloid precursor protein depends on lipid rafts. *J Cell Biol* **160**: 113-123.
104. Vetrivel, K. S., H. Cheng, W. Lin, T. Sakurai, T. Li, N. Nukina, P. C. Wong, H. Xu, and G. Thinakaran. 2004. Association of gamma-secretase with lipid rafts in post-Golgi and endosome membranes. *J Biol Chem* **279**: 44945-44954.
105. Lingwood, D., and K. Simons. 2010. Lipid rafts as a membrane-organizing principle. *Science* **327**: 46-50.
106. Simons, K., and J. L. Sampaio. 2011. Membrane organization and lipid rafts. *Cold Spring Harb Perspect Biol* **3**: a004697.

107. Cossec, J. C., A. Simon, C. Marquer, R. X. Moldrich, C. Leterrier, J. Rossier, C. Duyckaerts, Z. Lenkei, and M. C. Potier. 2010. Clathrin-dependent APP endocytosis and Abeta secretion are highly sensitive to the level of plasma membrane cholesterol. *Biochim Biophys Acta* **1801**: 846-852.
108. Yanagisawa, K. 2015. GM1 ganglioside and Alzheimer's disease. *Glycoconj J* **32**: 87-91.
109. Belkouch, M., M. Hachem, A. Elgot, A. Lo Van, M. Picq, M. Guichardant, M. Lagarde, and N. Bernoud-Hubac. 2016. The pleiotropic effects of omega-3 docosahexaenoic acid on the hallmarks of Alzheimer's disease. *J Nutr Biochem* **38**: 1-11.
110. Yassine, H. N., M. N. Braskie, W. J. Mack, K. J. Castor, A. N. Fonteh, L. S. Schneider, M. G. Harrington, and H. C. Chui. 2017. Association of Docosahexaenoic Acid Supplementation With Alzheimer Disease Stage in Apolipoprotein E epsilon4 Carriers: A Review. *JAMA Neurol*.
111. Grimm, M. O., H. S. Grimm, I. Tomic, K. Beyreuther, T. Hartmann, and C. Bergmann. 2008. Independent inhibition of Alzheimer disease beta- and gamma-secretase cleavage by lowered cholesterol levels. *J Biol Chem* **283**: 11302-11311.
112. Burg, V. K., H. S. Grimm, T. L. Rothhaar, S. Grosgen, B. Hundsdorfer, V. J. Haupenthal, V. C. Zimmer, J. Mett, O. Weingartner, U. Laufs, L. M. Broersen, H. Tanila, T. Vanmierlo, D. Lutjohann, T. Hartmann, and M. O. Grimm. 2013. Plant sterols the better cholesterol in Alzheimer's disease? A mechanistical study. *J Neurosci* **33**: 16072-16087.
113. Rothhaar, T. L., S. Grosgen, V. J. Haupenthal, V. K. Burg, B. Hundsdorfer, J. Mett, M. Riemenschneider, H. S. Grimm, T. Hartmann, and M. O. Grimm. 2012. Plasmalogens inhibit APP processing by directly affecting gamma-secretase activity in Alzheimer's disease. *ScientificWorldJournal* **2012**: 141240.
114. Grimm, M. O., V. J. Haupenthal, T. L. Rothhaar, V. C. Zimmer, S. Grosgen, B. Hundsdorfer, J. Lehmann, H. S. Grimm, and T. Hartmann. 2013. Effect of Different Phospholipids on alpha-Secretase Activity in the Non-Amyloidogenic Pathway of Alzheimer's Disease. *Int J Mol Sci* **14**: 5879-5898.
115. Koivisto, H., M. O. Grimm, T. L. Rothhaar, R. Berkecz, D. D. Lutjohann, R. Giniatullina, M. Takalo, P. O. Miettinen, H. M. Lahtinen, R. Giniatullin, B. Penke, T. Janaky, L. M. Broersen, T. Hartmann, and H. Tanila. 2014. Special lipid-based diets alleviate cognitive deficits in the

APPswe/PS1dE9 transgenic mouse model of Alzheimer's disease independent of brain amyloid deposition. *J Nutr Biochem* **25**: 157-169.

116. Karaca, I., I. Y. Tamboli, K. Glebov, J. Richter, L. H. Fell, M. O. Grimm, V. J. Haupenthal, T. Hartmann, M. H. Graler, G. van Echten-Deckert, and J. Walter. 2014. Deficiency of sphingosine-1-phosphate lyase impairs lysosomal metabolism of the amyloid precursor protein. *J Biol Chem* **289**: 16761-16772.

117. Grimm, M. O., T. L. Rothhaar, S. Grosgen, V. K. Burg, B. Hundsdorfer, V. J. Haupenthal, P. Friess, S. Kins, H. S. Grimm, and T. Hartmann. 2012. Trans fatty acids enhance amyloidogenic processing of the Alzheimer amyloid precursor protein (APP). *J Nutr Biochem* **23**: 1214-1223.

118. Naudi, A., R. Cabre, M. Jove, V. Ayala, H. Gonzalo, M. Portero-Otin, I. Ferrer, and R. Pamplona. 2015. Lipidomics of human brain aging and Alzheimer's disease pathology. *Int Rev Neurobiol* **122**: 133-189.

119. Wood, P. L., B. L. Barnette, J. A. Kaye, J. F. Quinn, and R. L. Woltjer. 2015. Non-targeted lipidomics of CSF and frontal cortex grey and white matter in control, mild cognitive impairment, and Alzheimer's disease subjects. *Acta Neuropsychiatr* **27**: 270-278.

120. Kosicek, M., and S. Hecimovic. 2013. Phospholipids and Alzheimer's disease: alterations, mechanisms and potential biomarkers. *Int J Mol Sci* **14**: 1310-1322.

121. Grimm, M. O., S. Grosgen, M. Riemenschneider, H. Tanila, H. S. Grimm, and T. Hartmann. 2011. From brain to food: analysis of phosphatidylcholins, lyso-phosphatidylcholins and phosphatidylcholin-plasmalogens derivatives in Alzheimer's disease human post mortem brains and mice model via mass spectrometry. *J Chromatogr A* **1218**: 7713-7722.

122. Lichtenthaler, S. F. 2012. Alpha-secretase cleavage of the amyloid precursor protein: proteolysis regulated by signaling pathways and protein trafficking. *Curr Alzheimer Res* **9**: 165-177.

123. Vingtdeux, V., and P. Marambaud. 2012. Identification and biology of alpha-secretase. *J Neurochem* **120 Suppl 1**: 34-45.

124. Buxbaum, J. D., K. N. Liu, Y. Luo, J. L. Slack, K. L. Stocking, J. J. Peschon, R. S. Johnson, B. J. Castner, D. P. Cerretti, and R. A. Black. 1998. Evidence that tumor necrosis factor alpha converting

enzyme is involved in regulated alpha-secretase cleavage of the Alzheimer amyloid protein precursor. *J Biol Chem* **273**: 27765-27767.

125. Koike, H., S. Tomioka, H. Sorimachi, T. C. Saido, K. Maruyama, A. Okuyama, A. Fujisawa-Sehara, S. Ohno, K. Suzuki, and S. Ishiura. 1999. Membrane-anchored metalloprotease MDC9 has an alpha-secretase activity responsible for processing the amyloid precursor protein. *Biochem J* **343 Pt 2**: 371-375.

126. Lammich, S., E. Kojro, R. Postina, S. Gilbert, R. Pfeiffer, M. Jasionowski, C. Haass, and F. Fahrenholz. 1999. Constitutive and regulated alpha-secretase cleavage of Alzheimer's amyloid precursor protein by a disintegrin metalloprotease. *Proc Natl Acad Sci U S A* **96**: 3922-3927.

127. Haass, C., A. Y. Hung, M. G. Schlossmacher, D. B. Teplow, and D. J. Selkoe. 1993. beta-Amyloid peptide and a 3-kDa fragment are derived by distinct cellular mechanisms. *J Biol Chem* **268**: 3021-3024.

128. Parr-Sturgess, C. A., D. J. Rushton, and E. T. Parkin. 2010. Ectodomain shedding of the Notch ligand Jagged1 is mediated by ADAM17, but is not a lipid-raft-associated event. *Biochem J* **432**: 283-294.

129. Bernardo, A., F. E. Harrison, M. McCord, J. Zhao, A. Bruchey, S. S. Davies, L. Jackson Roberts, 2nd, P. M. Mathews, Y. Matsuoka, T. Ariga, R. K. Yu, R. Thompson, and M. P. McDonald. 2009. Elimination of GD3 synthase improves memory and reduces amyloid-beta plaque load in transgenic mice. *Neurobiol Aging* **30**: 1777-1791.

130. Grimm, M. O. W., J. Mett, H. S. Grimm, and T. Hartmann. 2017. APP Function and Lipids: A Bidirectional Link. *Frontiers in Molecular Neuroscience* **10**.

131. Grimm, M. O., H. S. Grimm, A. J. Patzold, E. G. Zinser, R. Halonen, M. Duering, J. A. Tschape, B. De Strooper, U. Muller, J. Shen, and T. Hartmann. 2005. Regulation of cholesterol and sphingomyelin metabolism by amyloid-beta and presenilin. *Nat Cell Biol* **7**: 1118-1123.

132. Beel, A. J., C. K. Mobley, H. J. Kim, F. Tian, A. Hadziselimovic, B. Jap, J. H. Prestegard, and C. R. Sanders. 2008. Structural studies of the transmembrane C-terminal domain of the amyloid precursor protein (APP): does APP function as a cholesterol sensor? *Biochemistry* **47**: 9428-9446.

133. Sakurai, T., K. Kaneko, M. Okuno, K. Wada, T. Kashiya, H. Shimizu, T. Akagi, T. Hashikawa, and N. Nukina. 2008. Membrane microdomain switching: a regulatory mechanism of amyloid precursor protein processing. *J Cell Biol* **183**: 339-352.
134. He, X., Y. Huang, B. Li, C. X. Gong, and E. H. Schuchman. 2010. Deregulation of sphingolipid metabolism in Alzheimer's disease. *Neurobiol Aging* **31**: 398-408.
135. Alessenko, A. V., A. E. Bugrova, and L. B. Dudnik. 2004. Connection of lipid peroxide oxidation with the sphingomyelin pathway in the development of Alzheimer's disease. *Biochem Soc Trans* **32**: 144-146.
136. Grimm, M. O., J. Mett, C. P. Stahlmann, V. J. Haupenthal, T. Blumel, H. Stotzel, H. S. Grimm, and T. Hartmann. 2016. Eicosapentaenoic acid and docosahexaenoic acid increase the degradation of amyloid-beta by affecting insulin-degrading enzyme. *Biochem Cell Biol* **94**: 534-542.
137. Jana, A., and K. Pahan. 2004. Fibrillar amyloid-beta peptides kill human primary neurons via NADPH oxidase-mediated activation of neutral sphingomyelinase. Implications for Alzheimer's disease. *J Biol Chem* **279**: 51451-51459.
138. Jana, A., and K. Pahan. 2010. Fibrillar amyloid-beta-activated human astroglia kill primary human neurons via neutral sphingomyelinase: implications for Alzheimer's disease. *J Neurosci* **30**: 12676-12689.
139. Lee, J. K., H. K. Jin, M. H. Park, B. R. Kim, P. H. Lee, H. Nakauchi, J. E. Carter, X. He, E. H. Schuchman, and J. S. Bae. 2014. Acid sphingomyelinase modulates the autophagic process by controlling lysosomal biogenesis in Alzheimer's disease. *J Exp Med* **211**: 1551-1570.
140. Dinkins, M. B., J. Enasko, C. Hernandez, G. Wang, J. Kong, I. Helwa, Y. Liu, A. V. Terry, Jr., and E. Bieberich. 2016. Neutral Sphingomyelinase-2 Deficiency Ameliorates Alzheimer's Disease Pathology and Improves Cognition in the 5XFAD Mouse. *J Neurosci* **36**: 8653-8667.
141. Kornhuber, J., A. Medlin, S. Bleich, V. Jendrosseck, A. W. Henkel, J. Wiltfang, and E. Gulbins. 2005. High activity of acid sphingomyelinase in major depression. *J Neural Transm (Vienna)* **112**: 1583-1590.

142. Gao, Y., C. Huang, K. Zhao, L. Ma, X. Qiu, L. Zhang, Y. Xiu, L. Chen, W. Lu, C. Huang, Y. Tang, and Q. Xiao. 2013. Depression as a risk factor for dementia and mild cognitive impairment: a meta-analysis of longitudinal studies. *Int J Geriatr Psychiatry* **28**: 441-449.
143. Ellis, B., A. Hye, and S. G. Snowden. 2015. Metabolic Modifications in Human Biofluids Suggest the Involvement of Sphingolipid, Antioxidant, and Glutamate Metabolism in Alzheimer's Disease Pathogenesis. *J Alzheimers Dis* **46**: 313-327.
144. Koal, T., K. Klavins, D. Seppi, G. Kemmler, and C. Humpel. 2015. Sphingomyelin SM(d18:1/18:0) is significantly enhanced in cerebrospinal fluid samples dichotomized by pathological amyloid-beta42, tau, and phospho-tau-181 levels. *J Alzheimers Dis* **44**: 1193-1201.
145. Kosicek, M., H. Zetterberg, N. Andreasen, J. Peter-Katalinic, and S. Hecimovic. 2012. Elevated cerebrospinal fluid sphingomyelin levels in prodromal Alzheimer's disease. *Neurosci Lett* **516**: 302-305.
146. Olazaran, J., L. Gil-de-Gomez, A. Rodriguez-Martin, M. Valenti-Soler, B. Frades-Payo, J. Marin-Munoz, C. Antunez, A. Frank-Garcia, C. Acedo-Jimenez, L. Morlan-Gracia, R. Petidier-Torregrossa, M. C. Guisasola, F. Bermejo-Pareja, A. Sanchez-Ferro, D. A. Perez-Martinez, S. Manzano-Palomo, R. Farquhar, A. Rabano, and M. Calero. 2015. A blood-based, 7-metabolite signature for the early diagnosis of Alzheimer's disease. *J Alzheimers Dis* **45**: 1157-1173.
147. Gonzalez-Dominguez, R., T. Garcia-Barrera, J. Vitorica, and J. L. Gomez-Ariza. 2014. Region-specific metabolic alterations in the brain of the APP/PS1 transgenic mice of Alzheimer's disease. *Biochim Biophys Acta* **1842**: 2395-2402.
148. Katsel, P., C. Li, and V. Haroutunian. 2007. Gene expression alterations in the sphingolipid metabolism pathways during progression of dementia and Alzheimer's disease: a shift toward ceramide accumulation at the earliest recognizable stages of Alzheimer's disease? *Neurochem Res* **32**: 845-856.
149. Cutler, R. G., J. Kelly, K. Storie, W. A. Pedersen, A. Tammara, K. Hatanpaa, J. C. Troncoso, and M. P. Mattson. 2004. Involvement of oxidative stress-induced abnormalities in ceramide and

- cholesterol metabolism in brain aging and Alzheimer's disease. *Proc Natl Acad Sci U S A* **101**: 2070-2075.
150. Filippov, V., M. A. Song, K. Zhang, H. V. Vinters, S. Tung, W. M. Kirsch, J. Yang, and P. J. Duerksen-Hughes. 2012. Increased ceramide in brains with Alzheimer's and other neurodegenerative diseases. *J Alzheimers Dis* **29**: 537-547.
151. Han, X., S. Rozen, S. H. Boyle, C. Hellegers, H. Cheng, J. R. Burke, K. A. Welsh-Bohmer, P. M. Doraiswamy, and R. Kaddurah-Daouk. 2011. Metabolomics in early Alzheimer's disease: identification of altered plasma sphingolipidome using shotgun lipidomics. *PLoS One* **6**: e21643.
152. Mielke, M. M., V. V. Bandaru, N. J. Haughey, J. Xia, L. P. Fried, S. Yasar, M. Albert, V. Varma, G. Harris, E. B. Schneider, P. V. Rabins, K. Bandeen-Roche, C. G. Lyketsos, and M. C. Carlson. 2012. Serum ceramides increase the risk of Alzheimer disease: the Women's Health and Aging Study II. *Neurology* **79**: 633-641.
153. Couttas, T. A., N. Kain, A. K. Suchowerska, L. E. Quek, N. Turner, T. Fath, B. Garner, and A. S. Don. 2016. Loss of ceramide synthase 2 activity, necessary for myelin biosynthesis, precedes tau pathology in the cortical pathogenesis of Alzheimer's disease. *Neurobiol Aging* **43**: 89-100.
154. Mizuno, S., S. Ogishima, K. Kitatani, M. Kikuchi, H. Tanaka, N. Yaegashi, and J. Nakaya. 2016. Network Analysis of a Comprehensive Knowledge Repository Reveals a Dual Role for Ceramide in Alzheimer's Disease. *PLoS One* **11**: e0148431.
155. Puglielli, L., B. C. Ellis, A. J. Saunders, and D. M. Kovacs. 2003. Ceramide stabilizes beta-site amyloid precursor protein-cleaving enzyme 1 and promotes amyloid beta-peptide biogenesis. *J Biol Chem* **278**: 19777-19783.
156. Jazvinscak Jembrek, M., P. R. Hof, and G. Simic. 2015. Ceramides in Alzheimer's Disease: Key Mediators of Neuronal Apoptosis Induced by Oxidative Stress and Abeta Accumulation. *Oxid Med Cell Longev* **2015**: 346783.
157. van Echten-Deckert, G., N. Hagen-Euteneuer, I. Karaca, and J. Walter. 2014. Sphingosine-1-phosphate: boon and bane for the brain. *Cell Physiol Biochem* **34**: 148-157.

158. Takasugi, N., T. Sasaki, K. Suzuki, S. Osawa, H. Isshiki, Y. Hori, N. Shimada, T. Higo, S. Yokoshima, T. Fukuyama, V. M. Lee, J. Q. Trojanowski, T. Tomita, and T. Iwatsubo. 2011. BACE1 activity is modulated by cell-associated sphingosine-1-phosphate. *J Neurosci* **31**: 6850-6857.
159. Hagen, N., M. Hans, D. Hartmann, D. Swandulla, and G. van Echten-Deckert. 2011. Sphingosine-1-phosphate links glycosphingolipid metabolism to neurodegeneration via a calpain-mediated mechanism. *Cell Death Differ* **18**: 1356-1365.
160. Czubowicz, K., M. Cieslik, J. Pyszko, J. B. Strosznajder, and R. P. Strosznajder. 2015. Sphingosine-1-phosphate and its effect on glucose deprivation/glucose reload stress: from gene expression to neuronal survival. *Mol Neurobiol* **51**: 1300-1308.
161. Aytan, N., J. K. Choi, I. Carreras, V. Brinkmann, N. W. Kowall, B. G. Jenkins, and A. Dedeoglu. 2016. Fingolimod modulates multiple neuroinflammatory markers in a mouse model of Alzheimer's disease. *Sci Rep* **6**: 24939.
162. Marcus, J., S. Honigbaum, S. Shroff, K. Honke, J. Rosenbluth, and J. L. Dupree. 2006. Sulfatide is essential for the maintenance of CNS myelin and axon structure. *Glia* **53**: 372-381.
163. Han, X., M. H. D, D. W. McKeel, Jr., J. Kelley, and J. C. Morris. 2002. Substantial sulfatide deficiency and ceramide elevation in very early Alzheimer's disease: potential role in disease pathogenesis. *J Neurochem* **82**: 809-818.
164. Cheng, H., J. Xu, D. W. McKeel, Jr., and X. Han. 2003. Specificity and potential mechanism of sulfatide deficiency in Alzheimer's disease: an electrospray ionization mass spectrometric study. *Cell Mol Biol (Noisy-le-grand)* **49**: 809-818.
165. Cheng, H., Y. Zhou, D. M. Holtzman, and X. Han. 2010. Apolipoprotein E mediates sulfatide depletion in animal models of Alzheimer's disease. *Neurobiol Aging* **31**: 1188-1196.
166. Cheng, H., M. Wang, J. L. Li, N. J. Cairns, and X. Han. 2013. Specific changes of sulfatide levels in individuals with pre-clinical Alzheimer's disease: an early event in disease pathogenesis. *J Neurochem* **127**: 733-738.

167. Ariga, T., Y. Itokazu, M. P. McDonald, Y. Hirabayashi, S. Ando, and R. K. Yu. 2013. Brain gangliosides of a transgenic mouse model of Alzheimer's disease with deficiency in GD3-synthase: expression of elevated levels of a cholinergic-specific ganglioside, GT1aalpha. *ASN Neuro* **5**: 141-148.
168. Knight, E. M., H. N. Williams, A. C. Stevens, S. H. Kim, J. C. Kottwitz, A. D. Morant, J. W. Steele, W. L. Klein, K. Yanagisawa, R. E. Boyd, D. J. Lockhart, E. R. Sjoberg, M. E. Ehrlich, B. A. Wustman, and S. Gandy. 2015. Evidence that small molecule enhancement of beta-hexosaminidase activity corrects the behavioral phenotype in Dutch APP(E693Q) mice through reduction of ganglioside-bound Abeta. *Mol Psychiatry* **20**: 109-117.
169. Magini, A., A. Polchi, A. Tozzi, B. Tancini, M. Tantucci, L. Urbanelli, T. Borsello, P. Calabresi, and C. Emiliani. 2015. Abnormal cortical lysosomal beta-hexosaminidase and beta-galactosidase activity at post-synaptic sites during Alzheimer's disease progression. *Int J Biochem Cell Biol* **58**: 62-70.
170. Goetzl, E. J., A. Boxer, J. B. Schwartz, E. L. Abner, R. C. Petersen, B. L. Miller, and D. Kapogiannis. 2015. Altered lysosomal proteins in neural-derived plasma exosomes in preclinical Alzheimer disease. *Neurology* **85**: 40-47.
171. Perez, S. E., B. He, M. Nadeem, J. Wu, S. D. Ginsberg, M. D. Ikonovic, and E. J. Mufson. 2015. Hippocampal endosomal, lysosomal, and autophagic dysregulation in mild cognitive impairment: correlation with abeta and tau pathology. *J Neuropathol Exp Neurol* **74**: 345-358.
172. Zha, Q., Y. Ruan, T. Hartmann, K. Beyreuther, and D. Zhang. 2004. GM1 ganglioside regulates the proteolysis of amyloid precursor protein. *Mol Psychiatry* **9**: 946-952.
173. Kaya, I., D. Brinet, W. Michno, S. Syvanen, D. Sehlin, H. Zetterberg, K. Blennow, and J. Hanrieder. 2017. Delineating Amyloid Plaque Associated Neuronal Sphingolipids in Transgenic Alzheimer's Disease Mice (tgArcSwe) Using MALDI Imaging Mass Spectrometry. *ACS Chem Neurosci*.
174. Wakabayashi, M., T. Okada, Y. Kozutsumi, and K. Matsuzaki. 2005. GM1 ganglioside-mediated accumulation of amyloid beta-protein on cell membranes. *Biochem Biophys Res Commun* **328**: 1019-1023.

175. Ogawa, M., M. Tsukuda, T. Yamaguchi, K. Ikeda, T. Okada, Y. Yano, M. Hoshino, and K. Matsuzaki. 2011. Ganglioside-mediated aggregation of amyloid beta-proteins (Abeta): comparison between Abeta-(1-42) and Abeta-(1-40). *J Neurochem* **116**: 851-857.
176. Okada, T., K. Ikeda, M. Wakabayashi, M. Ogawa, and K. Matsuzaki. 2008. Formation of toxic Abeta(1-40) fibrils on GM1 ganglioside-containing membranes mimicking lipid rafts: polymorphisms in Abeta(1-40) fibrils. *J Mol Biol* **382**: 1066-1074.
177. Okada, T., M. Wakabayashi, K. Ikeda, and K. Matsuzaki. 2007. Formation of toxic fibrils of Alzheimer's amyloid beta-protein-(1-40) by monosialoganglioside GM1, a neuronal membrane component. *J Mol Biol* **371**: 481-489.
178. Ariga, T., K. Kobayashi, A. Hasegawa, M. Kiso, H. Ishida, and T. Miyatake. 2001. Characterization of high-affinity binding between gangliosides and amyloid beta-protein. *Arch Biochem Biophys* **388**: 225-230.
179. Molander-Melin, M., K. Blennow, N. Bogdanovic, B. Dellheden, J. E. Mansson, and P. Fredman. 2005. Structural membrane alterations in Alzheimer brains found to be associated with regional disease development; increased density of gangliosides GM1 and GM2 and loss of cholesterol in detergent-resistant membrane domains. *J Neurochem* **92**: 171-182.
180. Yamamoto, N., T. Matsubara, T. Sato, and K. Yanagisawa. 2008. Age-dependent high-density clustering of GM1 ganglioside at presynaptic neuritic terminals promotes amyloid beta-protein fibrillogenesis. *Biochim Biophys Acta* **1778**: 2717-2726.
181. Yanagisawa, K. 2011. Pathological significance of ganglioside clusters in Alzheimer's disease. *J Neurochem* **116**: 806-812.
182. Mutoh, T., Y. Hirabayashi, T. Mihara, M. Ueda, H. Koga, A. Ueda, T. Kokura, and H. Yamamoto. 2006. Role of glycosphingolipids and therapeutic perspectives on Alzheimer's disease. *CNS Neurol Disord Drug Targets* **5**: 375-380.
183. Marin, R., N. Fabelo, V. Martin, P. Garcia-Esparcia, I. Ferrer, D. Quinto-Aleman, and M. Diaz. 2017. Anomalies occurring in lipid profiles and protein distribution in frontal cortex lipid rafts in

dementia with Lewy bodies disclose neurochemical traits partially shared by Alzheimer's and Parkinson's diseases. *Neurobiol Aging* **49**: 52-59.

184. Chatterjee, P., W. L. Lim, G. Shui, V. B. Gupta, I. James, A. M. Fagan, C. Xiong, H. R. Sohrabi, K. Taddei, B. M. Brown, T. Benzinger, C. Masters, S. G. Snowden, M. R. Wenk, R. J. Bateman, J. C. Morris, and R. N. Martins. 2016. Plasma Phospholipid and Sphingolipid Alterations in Presenilin1 Mutation Carriers: A Pilot Study. *J Alzheimers Dis* **50**: 887-894.

185. Cheng, S. V., J. H. Nadeau, R. E. Tanzi, P. C. Watkins, J. Jagadesh, B. A. Taylor, J. L. Haines, N. Sacchi, and J. F. Gusella. 1988. Comparative mapping of DNA markers from the familial Alzheimer disease and Down syndrome regions of human chromosome 21 to mouse chromosomes 16 and 17. *Proc Natl Acad Sci U S A* **85**: 6032-6036.

186. Bueno, A. A., A. Brand, M. M. Neville, C. Lehane, N. Brierley, and M. A. Crawford. 2015. Erythrocyte phospholipid molecular species and fatty acids of Down syndrome children compared with non-affected siblings. *Br J Nutr* **113**: 72-81.

187. Onodera, T., E. Futai, E. Kan, N. Abe, T. Uchida, Y. Kamio, and J. Kaneko. 2015. Phosphatidylethanolamine plasmalogen enhances the inhibiting effect of phosphatidylethanolamine on gamma-secretase activity. *J Biochem* **157**: 301-309.

188. Ring, S., S. W. Weyer, S. B. Kilian, E. Waldron, C. U. Pietrzik, M. A. Filippov, J. Herms, C. Buchholz, C. B. Eckman, M. Korte, D. P. Wolfer, and U. C. Muller. 2007. The secreted beta-amyloid precursor protein ectodomain APPs alpha is sufficient to rescue the anatomical, behavioral, and electrophysiological abnormalities of APP-deficient mice. *J Neurosci* **27**: 7817-7826.

189. Yamashita, S., S. Kanno, A. Honjo, Y. Otoki, K. Nakagawa, M. Kinoshita, and T. Miyazawa. 2016. Analysis of Plasmalogen Species in Foodstuffs. *Lipids* **51**: 199-210.

190. Svennerholm, L. 1968. Distribution and fatty acid composition of phosphoglycerides in normal human brain. *J Lipid Res* **9**: 570-579.

191. Vors, C., J. Allaire, J. Marin, M. C. Lepine, A. Charest, A. Tchernof, P. Couture, and B. Lamarche. 2017. Inflammatory gene expression in whole blood cells after EPA vs. DHA supplementation: Results from the ComparED study. *Atherosclerosis* **257**: 116-122.

192. Hopperton, K. E., M. O. Trepanier, V. Giuliano, and R. P. Bazinet. 2016. Brain omega-3 polyunsaturated fatty acids modulate microglia cell number and morphology in response to intracerebroventricular amyloid-beta 1-40 in mice. *J Neuroinflammation* **13**: 257.
193. Famenini, S., E. A. Rigali, H. M. Olivera-Perez, J. Dang, M. T. Chang, R. Halder, R. V. Rao, M. Pellegrini, V. Porter, D. Bredesen, and M. Fiala. 2017. Increased intermediate M1-M2 macrophage polarization and improved cognition in mild cognitive impairment patients on omega-3 supplementation. *FASEB J* **31**: 148-160.
194. Fiala, M., R. C. Halder, B. Sagong, O. Ross, J. Sayre, V. Porter, and D. E. Bredesen. 2015. omega-3 Supplementation increases amyloid-beta phagocytosis and resolvin D1 in patients with minor cognitive impairment. *FASEB J* **29**: 2681-2689.
195. Mizwicki, M. T., G. Liu, M. Fiala, L. Magpantay, J. Sayre, A. Siani, M. Mahanian, R. Weitzman, E. Y. Hayden, M. J. Rosenthal, I. Nemere, J. Ringman, and D. B. Teplow. 2013. 1alpha,25-dihydroxyvitamin D3 and resolvin D1 retune the balance between amyloid-beta phagocytosis and inflammation in Alzheimer's disease patients. *J Alzheimers Dis* **34**: 155-170.
196. Stark, D. T., and N. G. Bazan. 2011. Neuroprotectin D1 induces neuronal survival and downregulation of amyloidogenic processing in Alzheimer's disease cellular models. *Mol Neurobiol* **43**: 131-138.
197. Bazan, N. G. 2009. Neuroprotectin D1-mediated anti-inflammatory and survival signaling in stroke, retinal degenerations, and Alzheimer's disease. *J Lipid Res* **50 Suppl**: S400-405.
198. van de Rest, O., N. van der Zwaluw, A. T. Beekman, L. C. de Groot, and J. M. Geleijnse. 2010. The reliability of three depression rating scales in a general population of Dutch older persons. *Int J Geriatr Psychiatry* **25**: 998-1005.
199. Stough, C., L. Downey, B. Silber, J. Lloyd, C. Kure, K. Wesnes, and D. Camfield. 2012. The effects of 90-day supplementation with the omega-3 essential fatty acid docosahexaenoic acid (DHA) on cognitive function and visual acuity in a healthy aging population. *Neurobiol Aging* **33**: 824 e821-823.

200. Yang, X., W. Sheng, G. Y. Sun, and J. C. Lee. 2011. Effects of fatty acid unsaturation numbers on membrane fluidity and alpha-secretase-dependent amyloid precursor protein processing. *Neurochem Int* **58**: 321-329.
201. Grimm, M. O., J. Kuchenbecker, S. Grosgen, V. K. Burg, B. Hundsdorfer, T. L. Rothhaar, P. Friess, M. C. de Wilde, L. M. Broersen, B. Penke, M. Peter, L. Vigh, H. S. Grimm, and T. Hartmann. 2011. Docosahexaenoic acid reduces amyloid beta production via multiple pleiotropic mechanisms. *J Biol Chem* **286**: 14028-14039.
202. Igarashi, M., K. Ma, F. Gao, H. W. Kim, S. I. Rapoport, and J. S. Rao. 2011. Disturbed choline plasmalogen and phospholipid fatty acid concentrations in Alzheimer's disease prefrontal cortex. *J Alzheimers Dis* **24**: 507-517.
203. Morris, M. C., D. A. Evans, J. L. Bienias, C. C. Tangney, D. A. Bennett, N. Aggarwal, J. Schneider, and R. S. Wilson. 2003. Dietary fats and the risk of incident Alzheimer disease. *Arch Neurol* **60**: 194-200.
204. Morris, M. C., D. A. Evans, J. L. Bienias, C. C. Tangney, and R. S. Wilson. 2004. Dietary fat intake and 6-year cognitive change in an older biracial community population. *Neurology* **62**: 1573-1579.
205. Morris, M. C., D. A. Evans, C. C. Tangney, J. L. Bienias, J. A. Schneider, R. S. Wilson, and P. A. Scherr. 2006. Dietary copper and high saturated and trans fat intakes associated with cognitive decline. *Arch Neurol* **63**: 1085-1088.
206. Muskiet, F. A., S. A. van Goor, R. S. Kuipers, F. V. Velzing-Aarts, E. N. Smit, H. Bouwstra, D. A. Dijck-Brouwer, E. R. Boersma, and M. Hadders-Algra. 2006. Long-chain polyunsaturated fatty acids in maternal and infant nutrition. *Prostaglandins Leukot Essent Fatty Acids* **75**: 135-144.
207. Lauritzen, L., H. S. Hansen, M. H. Jorgensen, and K. F. Michaelsen. 2001. The essentiality of long chain n-3 fatty acids in relation to development and function of the brain and retina. *Prog Lipid Res* **40**: 1-94.

208. Pawlosky, R. J., J. R. Hibbeln, J. A. Novotny, and N. Salem, Jr. 2001. Physiological compartmental analysis of alpha-linolenic acid metabolism in adult humans. *J Lipid Res* **42**: 1257-1265.
209. Nguyen, L. N., D. Ma, G. Shui, P. Wong, A. Cazenave-Gassiot, X. Zhang, M. R. Wenk, E. L. Goh, and D. L. Silver. 2014. Mfsd2a is a transporter for the essential omega-3 fatty acid docosahexaenoic acid. *Nature* **509**: 503-506.
210. Ouellet, M., V. Emond, C. T. Chen, C. Julien, F. Bourasset, S. Oddo, F. LaFerla, R. P. Bazinet, and F. Calon. 2009. Diffusion of docosahexaenoic and eicosapentaenoic acids through the blood-brain barrier: An in situ cerebral perfusion study. *Neurochem Int* **55**: 476-482.
211. Yassine, H. N., V. Rawat, W. J. Mack, J. F. Quinn, K. Yurko-Mauro, E. Bailey-Hall, P. S. Aisen, H. C. Chui, and L. S. Schneider. 2016. The effect of APOE genotype on the delivery of DHA to cerebrospinal fluid in Alzheimer's disease. *Alzheimers Res Ther* **8**: 25.
212. Kuusisto, J., K. Koivisto, K. Kervinen, L. Mykkanen, E. L. Helkala, M. Vanhanen, T. Hanninen, K. Pyorala, Y. A. Kesaniemi, P. Riekkinen, and et al. 1994. Association of apolipoprotein E phenotypes with late onset Alzheimer's disease: population based study. *BMJ* **309**: 636-638.
213. Breitner, J. C., B. W. Wyse, J. C. Anthony, K. A. Welsh-Bohmer, D. C. Steffens, M. C. Norton, J. T. Tschanz, B. L. Plassman, M. R. Meyer, I. Skoog, and A. Khachaturian. 1999. APOE-epsilon4 count predicts age when prevalence of AD increases, then declines: the Cache County Study. *Neurology* **53**: 321-331.
214. Tiraboschi, P., L. A. Hansen, E. Masliah, M. Alford, L. J. Thal, and J. Corey-Bloom. 2004. Impact of APOE genotype on neuropathologic and neurochemical markers of Alzheimer disease. *Neurology* **62**: 1977-1983.
215. Yassine, H. N., Q. Feng, I. Azizkhanian, V. Rawat, K. Castor, A. N. Fonteh, M. G. Harrington, L. Zheng, B. R. Reed, C. DeCarli, W. J. Jagust, and H. C. Chui. 2016. Association of Serum Docosahexaenoic Acid With Cerebral Amyloidosis. *JAMA Neurol* **73**: 1208-1216.

216. Horrocks, L. A., and A. A. Farooqui. 2004. Docosahexaenoic acid in the diet: its importance in maintenance and restoration of neural membrane function. *Prostaglandins Leukot Essent Fatty Acids* **70**: 361-372.
217. Grbovic, O. M., P. M. Mathews, Y. Jiang, S. D. Schmidt, R. Dinakar, N. B. Summers-Terio, B. P. Ceresa, R. A. Nixon, and A. M. Cataldo. 2003. Rab5-stimulated up-regulation of the endocytic pathway increases intracellular beta-cleaved amyloid precursor protein carboxyl-terminal fragment levels and Abeta production. *J Biol Chem* **278**: 31261-31268.
218. Carey, R. M., B. A. Balcz, I. Lopez-Coviella, and B. E. Slack. 2005. Inhibition of dynamin-dependent endocytosis increases shedding of the amyloid precursor protein ectodomain and reduces generation of amyloid beta protein. *BMC Cell Biol* **6**: 30.
219. Rajendran, L., M. Honsho, T. R. Zahn, P. Keller, K. D. Geiger, P. Verkade, and K. Simons. 2006. Alzheimer's disease beta-amyloid peptides are released in association with exosomes. *Proc Natl Acad Sci U S A* **103**: 11172-11177.
220. Huse, J. T., D. S. Pijak, G. J. Leslie, V. M. Lee, and R. W. Doms. 2000. Maturation and endosomal targeting of beta-site amyloid precursor protein-cleaving enzyme. The Alzheimer's disease beta-secretase. *J Biol Chem* **275**: 33729-33737.
221. Duncan, R. E., A. El-Sohemy, and M. C. Archer. 2005. Regulation of HMG-CoA reductase in MCF-7 cells by genistein, EPA, and DHA, alone and in combination with mevastatin. *Cancer Lett* **224**: 221-228.
222. Farris, W., S. Mansourian, Y. Chang, L. Lindsley, E. A. Eckman, M. P. Frosch, C. B. Eckman, R. E. Tanzi, D. J. Selkoe, and S. Guenette. 2003. Insulin-degrading enzyme regulates the levels of insulin, amyloid beta-protein, and the beta-amyloid precursor protein intracellular domain in vivo. *Proc Natl Acad Sci U S A* **100**: 4162-4167.
223. Hashimoto, M., S. Hossain, M. Katakura, A. Al Mamun, and O. Shido. 2015. The binding of Abeta1-42 to lipid rafts of RBC is enhanced by dietary docosahexaenoic acid in rats: Implicates to Alzheimer's disease. *Biochim Biophys Acta* **1848**: 1402-1409.

224. Mita, T., T. Mayanagi, H. Ichijo, K. Fukumoto, K. Otsuka, A. Sakai, and K. Sobue. 2016. Docosahexaenoic Acid Promotes Axon Outgrowth by Translational Regulation of Tau and Collapsin Response Mediator Protein 2 Expression. *J Biol Chem* **291**: 4955-4965.
225. Qu, M. H., X. Yang, Y. Wang, Q. Tang, H. Han, J. Wang, G. D. Wang, C. Xue, and Z. Gao. 2016. Docosahexaenoic Acid-Phosphatidylcholine Improves Cognitive Deficits in an Abeta23-35-Induced Alzheimer's Disease Rat Model. *Curr Top Med Chem* **16**: 558-564.
226. Florent, S., C. Malaplate-Armand, I. Youssef, B. Kriem, V. Koziel, M. C. Escanye, A. Fifre, I. Sponne, B. Leininger-Muller, J. L. Olivier, T. Pillot, and T. Oster. 2006. Docosahexaenoic acid prevents neuronal apoptosis induced by soluble amyloid-beta oligomers. *J Neurochem* **96**: 385-395.
227. Hossain, S., M. Hashimoto, M. Katakura, K. Miwa, T. Shimada, and O. Shido. 2009. Mechanism of docosahexaenoic acid-induced inhibition of in vitro Abeta1-42 fibrillation and Abeta1-42-induced toxicity in SH-SY5Y cells. *J Neurochem* **111**: 568-579.
228. Hashimoto, M., H. M. Shahdat, M. Katakura, Y. Tanabe, S. Gamoh, K. Miwa, T. Shimada, and O. Shido. 2009. Effects of docosahexaenoic acid on in vitro amyloid beta peptide 25-35 fibrillation. *Biochim Biophys Acta* **1791**: 289-296.
229. Teng, E., K. Taylor, T. Bilousova, D. Weiland, T. Pham, X. Zuo, F. Yang, P. P. Chen, C. G. Glabe, A. Takacs, D. R. Hoffman, S. A. Frautschy, and G. M. Cole. 2015. Dietary DHA supplementation in an APP/PS1 transgenic rat model of AD reduces behavioral and Abeta pathology and modulates Abeta oligomerization. *Neurobiol Dis* **82**: 552-560.
230. de Urquiza, A. M., S. Liu, M. Sjoberg, R. H. Zetterstrom, W. Griffiths, J. Sjoval, and T. Perlmann. 2000. Docosahexaenoic acid, a ligand for the retinoid X receptor in mouse brain. *Science* **290**: 2140-2144.
231. Lengqvist, J., A. Mata De Urquiza, A. C. Bergman, T. M. Willson, J. Sjoval, T. Perlmann, and W. J. Griffiths. 2004. Polyunsaturated fatty acids including docosahexaenoic and arachidonic acid bind to the retinoid X receptor alpha ligand-binding domain. *Mol Cell Proteomics* **3**: 692-703.

232. Forman, B. M., J. Chen, and R. M. Evans. 1997. Hypolipidemic drugs, polyunsaturated fatty acids, and eicosanoids are ligands for peroxisome proliferator-activated receptors alpha and delta. *Proc Natl Acad Sci U S A* **94**: 4312-4317.
233. Shysh, A. M., V. S. Nagibin, S. P. Kaplinskii, and V. E. Dosenko. 2016. N-3 long chain polyunsaturated fatty acids increase the expression of PPARgamma-target genes and resistance of isolated heart and cultured cardiomyocytes to ischemic injury. *Pharmacol Rep* **68**: 1133-1139.
234. Calder, P. C. 2015. Marine omega-3 fatty acids and inflammatory processes: Effects, mechanisms and clinical relevance. *Biochim Biophys Acta* **1851**: 469-484.
235. Casali, B. T., A. W. Corona, M. M. Mariani, J. C. Karlo, K. Ghosal, and G. E. Landreth. 2015. Omega-3 Fatty Acids Augment the Actions of Nuclear Receptor Agonists in a Mouse Model of Alzheimer's Disease. *J Neurosci* **35**: 9173-9181.
236. Song, C., C. H. Shieh, Y. S. Wu, A. Kalueff, S. Gaikwad, and K. P. Su. 2016. The role of omega-3 polyunsaturated fatty acids eicosapentaenoic and docosahexaenoic acids in the treatment of major depression and Alzheimer's disease: Acting separately or synergistically? *Prog Lipid Res* **62**: 41-54.
237. Bascul-Colombo, C., I. A. Guschina, B. H. Maskrey, M. Good, V. B. O'Donnell, and J. L. Harwood. 2016. Dietary DHA supplementation causes selective changes in phospholipids from different brain regions in both wild type mice and the Tg2576 mouse model of Alzheimer's disease. *Biochim Biophys Acta* **1861**: 524-537.
238. Diaz, M., N. Fabelo, V. Casanas-Sanchez, R. Marin, T. Gomez, D. Quinto-Aleman, and J. A. Perez. 2016. Hippocampal Lipid Homeostasis in APP/PS1 Mice is Modulated by a Complex Interplay Between Dietary DHA and Estrogens: Relevance for Alzheimer's Disease. *J Alzheimers Dis* **49**: 459-481.
239. Grimm, M. O., V. J. Haupenthal, J. Mett, C. P. Stahlmann, T. Blumel, N. T. Mylonas, K. Endres, H. S. Grimm, and T. Hartmann. 2016. Oxidized Docosahexaenoic Acid Species and Lipid Peroxidation Products Increase Amyloidogenic Amyloid Precursor Protein Processing. *Neurodegener Dis* **16**: 44-54.
240. Mohaibes, R. J., M. A. Fiol-deRoque, M. Torres, M. Ordinas, D. J. Lopez, J. A. Castro, P. V. Escriba, and X. Busquets. 2017. The hydroxylated form of docosahexaenoic acid (DHA-H) modifies the

brain lipid composition in a model of Alzheimer's disease, improving behavioral motor function and survival. *Biochim Biophys Acta*.

241. Cai, Z., B. Zhao, and A. Ratka. 2011. Oxidative stress and beta-amyloid protein in Alzheimer's disease. *Neuromolecular Med* **13**: 223-250.
242. Kim, G. H., J. E. Kim, S. J. Rhie, and S. Yoon. 2015. The Role of Oxidative Stress in Neurodegenerative Diseases. *Exp Neurobiol* **24**: 325-340.
243. Apelt, J., M. Bigl, P. Wunderlich, and R. Schliebs. 2004. Aging-related increase in oxidative stress correlates with developmental pattern of beta-secretase activity and beta-amyloid plaque formation in transgenic Tg2576 mice with Alzheimer-like pathology. *Int J Dev Neurosci* **22**: 475-484.
244. Manczak, M., T. S. Anekonda, E. Henson, B. S. Park, J. Quinn, and P. H. Reddy. 2006. Mitochondria are a direct site of A beta accumulation in Alzheimer's disease neurons: implications for free radical generation and oxidative damage in disease progression. *Hum Mol Genet* **15**: 1437-1449.
245. Rogers, L. K., C. J. Valentine, and S. A. Keim. 2013. DHA supplementation: current implications in pregnancy and childhood. *Pharmacol Res* **70**: 13-19.
246. Yurko-Mauro, K., D. McCarthy, D. Rom, E. B. Nelson, A. S. Ryan, A. Blackwell, N. Salem, Jr., and M. Stedman. 2010. Beneficial effects of docosahexaenoic acid on cognition in age-related cognitive decline. *Alzheimer's & dementia : the journal of the Alzheimer's Association* **6**: 456-464.
247. Plassman, B. L., J. W. Williams, Jr., J. R. Burke, T. Holsinger, and S. Benjamin. 2010. Systematic review: factors associated with risk for and possible prevention of cognitive decline in later life. *Ann Intern Med* **153**: 182-193.
248. Moriguchi, T., R. S. Greiner, and N. Salem, Jr. 2000. Behavioral deficits associated with dietary induction of decreased brain docosahexaenoic acid concentration. *J Neurochem* **75**: 2563-2573.
249. Salem, N., Jr., T. Moriguchi, R. S. Greiner, K. McBride, A. Ahmad, J. N. Catalan, and B. Slotnick. 2001. Alterations in brain function after loss of docosahexaenoate due to dietary restriction of n-3 fatty acids. *J Mol Neurosci* **16**: 299-307; discussion 317-221.

250. Calon, F., G. P. Lim, F. Yang, T. Morihara, B. Teter, O. Ubeda, P. Rostaing, A. Triller, N. Salem, Jr., K. H. Ashe, S. A. Frautschy, and G. M. Cole. 2004. Docosahexaenoic acid protects from dendritic pathology in an Alzheimer's disease mouse model. *Neuron* **43**: 633-645.
251. Soderberg, M., C. Edlund, I. Alafuzoff, K. Kristensson, and G. Dallner. 1992. Lipid composition in different regions of the brain in Alzheimer's disease/senile dementia of Alzheimer's type. *J Neurochem* **59**: 1646-1653.
252. Tully, A. M., H. M. Roche, R. Doyle, C. Fallon, I. Bruce, B. Lawlor, D. Coakley, and M. J. Gibney. 2003. Low serum cholesteryl ester-docosahexaenoic acid levels in Alzheimer's disease: a case-control study. *Br J Nutr* **89**: 483-489.
253. Albanese, E., A. D. Dangour, R. Uauy, D. Acosta, M. Guerra, S. S. Guerra, Y. Huang, K. S. Jacob, J. L. de Rodriguez, L. H. Noriega, A. Salas, A. L. Sosa, R. M. Sousa, J. Williams, C. P. Ferri, and M. J. Prince. 2009. Dietary fish and meat intake and dementia in Latin America, China, and India: a 10/66 Dementia Research Group population-based study. *The American journal of clinical nutrition* **90**: 392-400.
254. van Gelder, B. M., M. Tijhuis, S. Kalmijn, and D. Kromhout. 2007. Fish consumption, n-3 fatty acids, and subsequent 5-y cognitive decline in elderly men: the Zutphen Elderly Study. *The American journal of clinical nutrition* **85**: 1142-1147.
255. Astarita, G., K. M. Jung, N. C. Berchtold, V. Q. Nguyen, D. L. Gillen, E. Head, C. W. Cotman, and D. Piomelli. 2010. Deficient liver biosynthesis of docosahexaenoic acid correlates with cognitive impairment in Alzheimer's disease. *PloS one* **5**: e12538.
256. van de Rest, O., Y. Wang, L. L. Barnes, C. Tangney, D. A. Bennett, and M. C. Morris. 2016. APOE epsilon4 and the associations of seafood and long-chain omega-3 fatty acids with cognitive decline. *Neurology* **86**: 2063-2070.
257. Zhang, Y., J. Chen, J. Qiu, Y. Li, J. Wang, and J. Jiao. 2016. Intakes of fish and polyunsaturated fatty acids and mild-to-severe cognitive impairment risks: a dose-response meta-analysis of 21 cohort studies. *Am J Clin Nutr* **103**: 330-340.

258. Freund-Levi, Y., M. Eriksdotter-Jonhagen, T. Cederholm, H. Basun, G. Faxen-Irving, A. Garlind, I. Vedin, B. Vessby, L. O. Wahlund, and J. Palmblad. 2006. Omega-3 fatty acid treatment in 174 patients with mild to moderate Alzheimer disease: OmegAD study: a randomized double-blind trial. *Arch Neurol* **63**: 1402-1408.
259. Quinn, J. F., R. Raman, R. G. Thomas, K. Yurko-Mauro, E. B. Nelson, C. Van Dyck, J. E. Galvin, J. Emond, C. R. Jack, Jr., M. Weiner, L. Shinto, and P. S. Aisen. 2010. Docosahexaenoic acid supplementation and cognitive decline in Alzheimer disease: a randomized trial. *JAMA* **304**: 1903-1911.
260. Frautschy, S. A., and G. M. Cole. 2011. What was lost in translation in the DHA trial is whom you should intend to treat. *Alzheimer's research & therapy* **3**: 2.
261. Smith, A. D., S. M. Smith, C. A. de Jager, P. Whitbread, C. Johnston, G. Agacinski, A. Oulhaj, K. M. Bradley, R. Jacoby, and H. Refsum. 2010. Homocysteine-lowering by B vitamins slows the rate of accelerated brain atrophy in mild cognitive impairment: a randomized controlled trial. *PLoS one* **5**: e12244.
262. de Jager, C. A., A. Oulhaj, R. Jacoby, H. Refsum, and A. D. Smith. 2012. Cognitive and clinical outcomes of homocysteine-lowering B-vitamin treatment in mild cognitive impairment: a randomized controlled trial. *Int J Geriatr Psychiatry* **27**: 592-600.
263. Seshadri, S., A. Beiser, J. Selhub, P. F. Jacques, I. H. Rosenberg, R. B. D'Agostino, P. W. Wilson, and P. A. Wolf. 2002. Plasma homocysteine as a risk factor for dementia and Alzheimer's disease. *N Engl J Med* **346**: 476-483.
264. Douaud, G., H. Refsum, C. A. de Jager, R. Jacoby, T. E. Nichols, S. M. Smith, and A. D. Smith. 2013. Preventing Alzheimer's disease-related gray matter atrophy by B-vitamin treatment. *Proc Natl Acad Sci U S A* **110**: 9523-9528.
265. Martins, J. G. 2009. EPA but not DHA appears to be responsible for the efficacy of omega-3 long chain polyunsaturated fatty acid supplementation in depression: evidence from a meta-analysis of randomized controlled trials. *J Am Coll Nutr* **28**: 525-542.

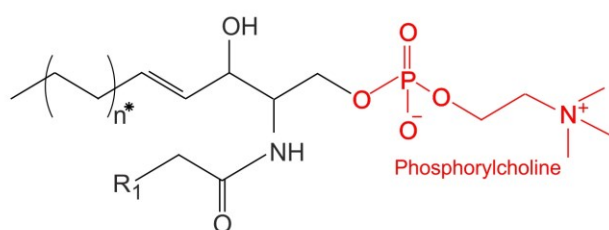
266. De Bruin, N. M., A. J. Kiliaan, M. C. De Wilde, and L. M. Broersen. 2003. Combined uridine and choline administration improves cognitive deficits in spontaneously hypertensive rats. *Neurobiol Learn Mem* **80**: 63-79.
267. Kryscio, R. J., E. L. Abner, F. A. Schmitt, P. J. Goodman, M. Mendiondo, A. Caban-Holt, B. C. Dennis, M. Mathews, E. A. Klein, and J. J. Crowley. 2013. A randomized controlled Alzheimer's disease prevention trial's evolution into an exposure trial: the PREADViSE Trial. *J Nutr Health Aging* **17**: 72-75.
268. Wurtman, R. J., I. H. Ulus, M. Cansev, C. J. Watkins, L. Wang, and G. Marzloff. 2006. Synaptic proteins and phospholipids are increased in gerbil brain by administering uridine plus docosahexaenoic acid orally. *Brain research* **1088**: 83-92.
269. Cansev, M., R. J. Wurtman, T. Sakamoto, and I. H. Ulus. 2008. Oral administration of circulating precursors for membrane phosphatides can promote the synthesis of new brain synapses. *Alzheimer's & dementia : the journal of the Alzheimer's Association* **4**: S153-168.
270. Lim, S. Y., and H. Suzuki. 2000. Intakes of dietary docosahexaenoic acid ethyl ester and egg phosphatidylcholine improve maze-learning ability in young and old mice. *J Nutr* **130**: 1629-1632.
271. de Wilde, M. C., E. Farkas, M. Gerrits, A. J. Kiliaan, and P. G. Luiten. 2002. The effect of n-3 polyunsaturated fatty acid-rich diets on cognitive and cerebrovascular parameters in chronic cerebral hypoperfusion. *Brain research* **947**: 166-173.
272. de Wilde, M. C., E. Högges, A. J. Kiliaan, T. Farkas, P. G. Luiten, and E. Farkas. 2003. Dietary fatty acids alter blood pressure, behavior and brain membrane composition of hypertensive rats. *Brain research* **988**: 9-19.
273. Savelkoul, P. J., H. Janickova, A. A. Kuipers, R. J. Hageman, P. J. Kamphuis, V. Dolezal, and L. M. Broersen. 2012. A specific multi-nutrient formulation enhances M1 muscarinic acetylcholine receptor responses in vitro. *J Neurochem* **120**: 631-640.
274. Jansen, D., V. Zerbi, I. A. Arnoldussen, M. Wiesmann, A. Rijpma, X. T. Fang, P. J. Dederen, M. P. Mutsaers, L. M. Broersen, D. Lutjohann, M. Miller, L. A. Joosten, A. Heerschap, and A. J. Kiliaan. 2013. Effects of Specific Multi-Nutrient Enriched Diets on Cerebral Metabolism, Cognition and Neuropathology in AbetaPPswe-PS1dE9 Mice. *PLoS one* **8**: e75393.

275. van Wijk, N., L. M. Broersen, M. C. de Wilde, R. J. Hageman, M. Groenendijk, J. W. Sijben, and P. J. Kamphuis. 2013. Targeting Synaptic Dysfunction in Alzheimer's Disease by Administering a Specific Nutrient Combination. *J Alzheimers Dis*.
276. Wurtman, R. J., M. Cansev, T. Sakamoto, and I. H. Ulus. 2009. Use of phosphatide precursors to promote synaptogenesis. *Annual review of nutrition* **29**: 59-87.
277. Broersen, L. M., A. A. Kuipers, M. Balvers, N. van Wijk, P. J. Savelkoul, M. C. de Wilde, E. M. van der Beek, J. W. Sijben, R. J. Hageman, P. J. Kamphuis, and A. J. Kiliaan. 2013. A specific multi-nutrient diet reduces Alzheimer-like pathology in young adult AbetaPPswe/PS1dE9 mice. *J Alzheimers Dis* **33**: 177-190.
278. Cansev, M., N. van Wijk, M. Turkeyilmaz, F. Orhan, J. W. Sijben, and L. M. Broersen. 2015. Specific multi-nutrient enriched diet enhances hippocampal cholinergic transmission in aged rats. *Neurobiol Aging* **36**: 344-351.
279. Janickova, H., V. Rudajev, E. Dolejsi, H. Koivisto, J. Jakubik, H. Tanila, E. E. El-Fakahany, and V. Dolezal. 2015. Lipid-Based Diets Improve Muscarinic Neurotransmission in the Hippocampus of Transgenic APPswe/PS1dE9 Mice. *Curr Alzheimer Res* **12**: 923-931.
280. Zerbi, V., D. Jansen, M. Wiesmann, X. Fang, L. M. Broersen, A. Veltien, A. Heerschap, and A. J. Kiliaan. 2014. Multinutrient diets improve cerebral perfusion and neuroprotection in a murine model of Alzheimer's disease. *Neurobiol Aging* **35**: 600-613.
281. Jansen, D., V. Zerbi, C. I. Janssen, D. van Rooij, B. Zinnhardt, P. J. Dederen, A. J. Wright, L. M. Broersen, D. Lutjohann, A. Heerschap, and A. J. Kiliaan. 2013. Impact of a multi-nutrient diet on cognition, brain metabolism, hemodynamics, and plasticity in apoE4 carrier and apoE knockout mice. *Brain Struct Funct*.
282. Scarmeas, N., Y. Stern, R. Mayeux, J. J. Manly, N. Schupf, and J. A. Luchsinger. 2009. Mediterranean diet and mild cognitive impairment. *Arch Neurol* **66**: 216-225.
283. Oulhaj, A., F. Jerneren, H. Refsum, A. D. Smith, and C. A. de Jager. 2016. Omega-3 Fatty Acid Status Enhances the Prevention of Cognitive Decline by B Vitamins in Mild Cognitive Impairment. *J Alzheimers Dis* **50**: 547-557.

284. Jerneren, F., A. K. Elshorbagy, A. Oulhaj, S. M. Smith, H. Refsum, and A. D. Smith. 2015. Brain atrophy in cognitively impaired elderly: the importance of long-chain omega-3 fatty acids and B vitamin status in a randomized controlled trial. *The American journal of clinical nutrition* **102**: 215-221.
285. Scheltens, P., P. J. Kamphuis, F. R. Verhey, M. G. Olde Rikkert, R. J. Wurtman, D. Wilkinson, J. W. Twisk, and A. Kurz. 2010. Efficacy of a medical food in mild Alzheimer's disease: A randomized, controlled trial. *Alzheimer's & dementia : the journal of the Alzheimer's Association* **6**: 1-10 e11.
286. Scheltens, P., J. W. Twisk, R. Blesa, E. Scarpini, C. A. von Arnim, A. Bongers, J. Harrison, S. H. Swinkels, C. J. Stam, H. de Waal, R. J. Wurtman, R. L. Wieggers, B. Vellas, and P. J. Kamphuis. 2012. Efficacy of Souvenaid in mild Alzheimer's disease: results from a randomized, controlled trial. *J Alzheimers Dis* **31**: 225-236.
287. de Waal, H., C. J. Stam, M. M. Lansbergen, R. L. Wieggers, P. J. Kamphuis, P. Scheltens, F. Maestu, and E. C. van Straaten. 2014. The effect of souvenaid on functional brain network organisation in patients with mild Alzheimer's disease: a randomised controlled study. *PloS one* **9**: e86558.
288. Shah, R. C., P. J. Kamphuis, S. Leurgans, S. H. Swinkels, C. H. Sadowsky, A. Bongers, S. A. Rappaport, J. F. Quinn, R. L. Wieggers, P. Scheltens, and D. A. Bennett. 2013. The S-Connect study: results from a randomized, controlled trial of Souvenaid in mild-to-moderate Alzheimer's disease. *Alzheimer's research & therapy* **5**: 59.
289. Cummings, J., P. Scheltens, I. McKeith, R. Blesa, J. E. Harrison, P. H. Bertolucci, K. Rockwood, D. Wilkinson, W. Wijker, D. A. Bennett, and R. C. Shah. 2017. Effect Size Analyses of Souvenaid in Patients with Alzheimer's Disease. *J Alzheimers Dis* **55**: 1131-1139.

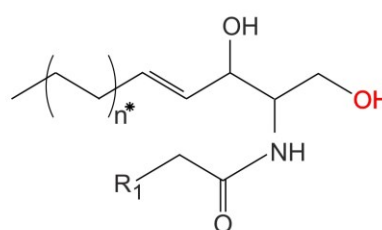
Figure 1:

Sphingomyelin (SM):

R₁ = different acyl chains

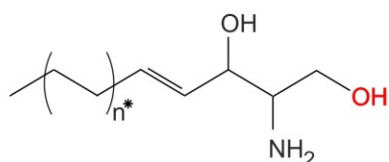
n* = in many cases 6

Ceramide (Cer):

R₁ = different acyl chains

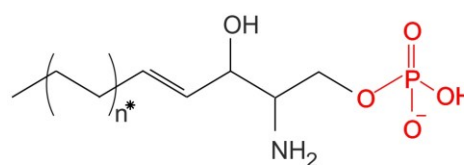
n* = in many cases 6

Sphingosine (Sph):



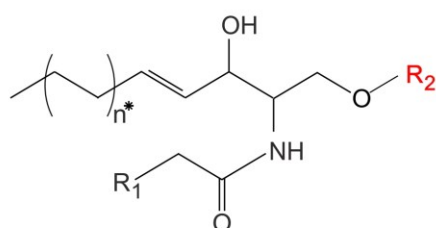
n* = in many cases 6

Sphingosine-1-Phosphate (S1P):



n* = in many cases 6

Gangliosides

R₁ = different acyl chains

n* = in many cases 6

R₂ = different carbohydrates containing one or more sialic acids and derivatives and different sugars

Figure 1: Chemical structure of the sphingolipids which are altered in AD.

Figure 2:

Plasmalogen (PL):

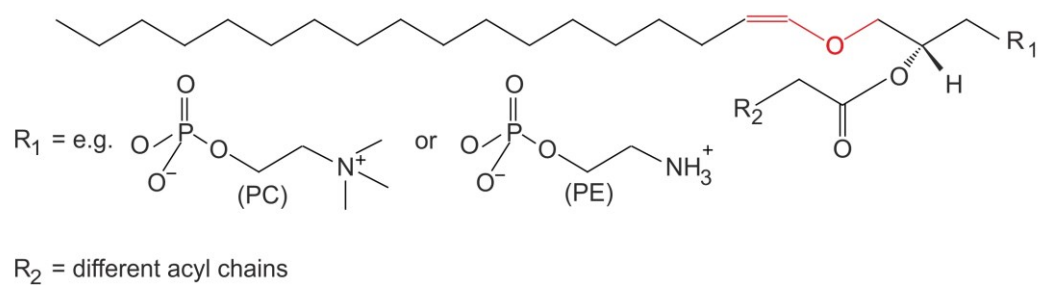
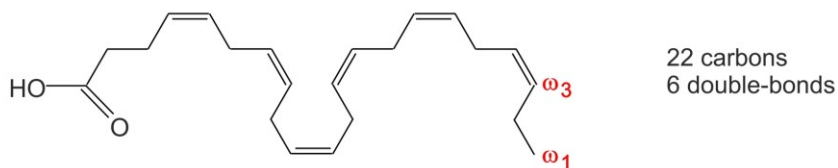


Figure 2: Chemical structure of plasmalogens

Figure 3:

DHA:



EPA:

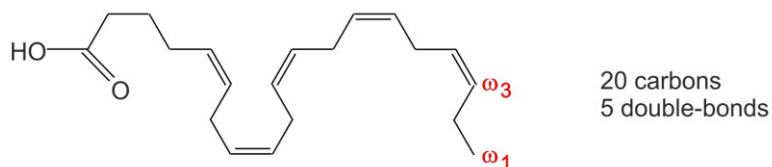


Figure 3: Chemical structure of the fatty acids DHA and EPA